



African Journal of Pharmacy and Pharmacology

Volume 9 Number 3, 22 January, 2015

ISSN 1996-0816



*Academic
Journals*

ABOUT AJPP

The **African Journal of Pharmacy and Pharmacology (AJPP)** is published weekly (one volume per year) by Academic Journals.

African Journal of Pharmacy and Pharmacology (AJPP) is an open access journal that provides rapid publication (weekly) of articles in all areas of Pharmaceutical Science such as Pharmaceutical Microbiology, Pharmaceutical Raw Material Science, Formulations, Molecular modeling, Health sector Reforms, Drug Delivery, Pharmacokinetics and Pharmacodynamics, Pharmacognosy, Social and Administrative Pharmacy, Pharmaceutics and Pharmaceutical Microbiology, Herbal Medicines research, Pharmaceutical Raw Materials development/utilization, Novel drug delivery systems, Polymer/Cosmetic Science, Food/Drug Interaction, Herbal drugs evaluation, Physical Pharmaceutics, Medication management, Cosmetic Science, pharmaceuticals, pharmacology, pharmaceutical research etc. The Journal welcomes the submission of manuscripts that meet the general criteria of significance and scientific excellence. Papers will be published shortly after acceptance. All articles published in AJPP are peer-reviewed.

Submission of Manuscript

Submit manuscripts as e-mail attachment to the Editorial Office at: ajpp@academicjournals.org. A manuscript number will be mailed to the corresponding author shortly after submission.

The African Journal of Pharmacy and Pharmacology will only accept manuscripts submitted as e-mail attachments.

Please read the **Instructions for Authors** before submitting your manuscript. The manuscript files should be given the last name of the first author.

Editors

Sharmilah Pamela Seetulsingh- Goorah

*Associate Professor,
Department of Health Sciences
Faculty of Science,
University of Mauritius,
Reduit,
Mauritius*

Himanshu Gupta

*University of Colorado- Anschutz Medical Campus,
Department of Pharmaceutical Sciences, School of
Pharmacy Aurora, CO 80045,
USA*

Dr. Shreesh Kumar Ojha

*Molecular Cardiovascular Research Program
College of Medicine
Arizona Health Sciences Center
University of Arizona
Tucson 85719, Arizona,
USA*

Dr. Victor Valenti Engracia

*Department of Speech-Language and
Hearing Therapy Faculty of Philosophy
and Sciences, UNESP
Marilia-SP, Brazil.*

Prof. Sutiak Vaclav

*Rovníková 7, 040 20 Košice,
The Slovak Republic,
The Central Europe,
European Union
Slovak Republic
Slovakia*

Dr.B.RAVISHANKAR

*Director and Professor of Experimental Medicine
SDM Centre for Ayurveda and Allied Sciences,
SDM College of Ayurveda Campus,
Kuthpady, Udupi- 574118
Karnataka (INDIA)*

Dr. Manal Moustafa Zaki

*Department of Veterinary Hygiene and Management
Faculty of Veterinary Medicine, Cairo University
Giza, 11221 Egypt*

Prof. George G. Nomikos

*Scientific Medical Director
Clinical Science
Neuroscience
TAKEDA GLOBAL RESEARCH & DEVELOPMENT
CENTER, INC. 675 North Field Drive Lake Forest, IL
60045
USA*

Prof. Mahmoud Mohamed El-Mas

Department of Pharmacology,

Dr. Caroline Wagner

*Universidade Federal do Pampa
Avenida Pedro Anunciação, s/n
Vila Batista, Caçapava do Sul, RS - Brazil*

Editorial Board

Prof. Fen Jicai

School of life science, Xinjiang University, China.

Dr. Ana Laura Nicoletti Carvalho

Av. Dr. Arnaldo, 455, São Paulo, SP. Brazil.

Dr. Ming-hui Zhao

*Professor of Medicine
Director of Renal Division, Department of Medicine
Peking University First Hospital
Beijing 100034
PR. China.*

Prof. Ji Junjun

Guangdong Cardiovascular Institute, Guangdong General Hospital, Guangdong Academy of Medical Sciences, China.

Prof. Yan Zhang

*Faculty of Engineering and Applied Science,
Memorial University of Newfoundland,
Canada.*

Dr. Naoufel Madani

*Medical Intensive Care Unit
University hospital Ibn Sina, Univesity Mohamed V
Souissi, Rabat,
Morocco.*

Dr. Dong Hui

Department of Gynaecology and Obstetrics, the 1st hospital, NanFang University, China.

Prof. Ma Hui

School of Medicine, Lanzhou University, China.

Prof. Gu HuiJun

School of Medicine, Taizhou university, China.

Dr. Chan Kim Wei

*Research Officer
Laboratory of Molecular Biomedicine,
Institute of Bioscience, Universiti Putra,
Malaysia.*

Dr. Fen Cun

Professor, Department of Pharmacology, Xinjiang University, China.

Dr. Sirajunnisa Razack

Department of Chemical Engineering, Annamalai University, Annamalai Nagar, Tamilnadu, India.

Prof. Ehab S. EL Desoky

Professor of pharmacology, Faculty of Medicine Assiut University, Assiut, Egypt.

Dr. Yakisich, J. Sebastian

Assistant Professor, Department of Clinical Neuroscience R54 Karolinska University Hospital, Huddinge 141 86 Stockholm , Sweden.

Prof. Dr. Andrei N. Tchernitchin

Head, Laboratory of Experimental Endocrinology and Environmental Pathology LEEPA University of Chile Medical School, Chile.

Dr. Sirajunnisa Razack

Department of Chemical Engineering, Annamalai University, Annamalai Nagar, Tamilnadu, India.

Dr. Yasar Tatar

Marmara University, Turkey.

Dr Nafisa Hassan Ali

Assistant Professor, Dow institute of medical technology Dow University of Health Sciences, Chand bbi Road, Karachi, Pakistan.

Dr. Krishnan Namboori P. K.

Computational Chemistry Group, Computational Engineering and Networking, Amrita Vishwa Vidyapeetham, Amritanagar, Coimbatore-641 112 India.

Prof. Osman Ghani

University of Sargodha, Pakistan.

Dr. Liu Xiaoji

School of Medicine, Shihezi University, China.

African Journal of Pharmacy and Pharmacology

Table of Contents: Volume 9 Number 3 22 January, 2015

ARTICLES

Research Articles

- A Review On Novel Therapies To Combat Hepatitis C** 37
Pragati Khare, Lubhan Singh and Ghanshyam Yadav
- Determination Of The Abortifacient Activity Of The Aqueous Extract Of Phytolacca Dodecandra (L'Her) Leaf In Wistar Rats** 43
Angella Namulindwa, David Nkwangu and Joseph Oloro
- Analysis Of Rational Use Of Drugs As Of Facility Indicators And Patient Care Indicators Practices At Four Selected Hospitals Of West Ethiopia: Policy Implication** 48
Tadesse Haile Fereja and Jimma Likisa Lenjesa
- Anxiolytic Properties Of Melissa Officinalis And Associated Mechanisms Of Action: A Review Of The Literature** 53
Bárbara Luisa Fermino, Najeh Nasser Kahlil, Juliana Sartori Bonini, Romaiana Picada Pereira, João Batista Teixeira da Rocha and Weber Claudio Francisco Nunes da Silva
- Effect Of Citrus Paradisi And Citrus Sinensis On Glycemic Control In Rats** 60
Neelam Mallick and Rafeeq Alam Khan

Full Length Research Paper

Exploration of the neurotoxicity of ciprofloxacin or gatifloxacin single dose in rat cortex and hippocampus

Nadia Mohamed Said Arafa^{1*}, Sayed M. Rawi² and Sara Abdullah Mubarak³

¹Department of Biology, Faculty of Science, Jazan University, KSA & National Organization for Drug Control and Research, Department of Physiology, Egypt.

²Department of Biology, Faculty of Sciences and Arts, Khulais, Jeddah University, Saudi Arabia.

³Department of Dairy Lab, Public Authority of Agriculture & Fish Resources (PAAFR), State of Kuwait.

Received 15 November, 2014; Accepted 29 January, 2015

The study aimed to evaluate the neurotoxicity of ciprofloxacin (Cip) or gatifloxacin (Gati) single oral dose in male albino rats weighing (100 ± 20 g) grouped as control-administered water, ciprofloxacin (80 mg/kg) and gatifloxacin (32 mg/kg) each of 12 rats. The frontal cortex of both groups revealed decrease in glutamate, GABA, taurine, histidine and serotonin levels and elevation of aspartate, glycin and serine and AChE activities. While noradrenaline and dopamine levels reduced significantly in Gati group, noradrenaline increased significantly in Cip group. Hippocampus of either Cip or Gati group's results revealed elevation of all detected amino acids and monoamines except the reduction of glutamate, aspartate and dopamine in Cip group. In the meantime, AChE activities significantly reduced in both treatments. Serum results showed elevation of glucose in both treated groups. The histological examination of Gati brain tissue showed neuronal degeneration in the cerebral cortex and congestion in the blood vessels and capillaries in hippocampus tissue without histopathological alteration observed in Cip group tissue. Overall, the data showed the effect of the quinolones single dose towards hyperglycemia and shift in balance of neurotransmitters and acetylcholinesterase as well as the histopathological alterations in the tested brain areas.

Key words: Ciprofloxacin, gatifloxacin, cortex, hippocampus, neurotransmitters, glucose.

INTRODUCTION

Gatifloxacin is one of the broad-spectrum fluoroquinolones available and approved by the US food and drug administration (FDA) in December 1999. Ever since its release in the market, there have been numerous reports implicating gatifloxacin as a cause of

hypoglycemia and hyperglycemia. This prompted Bristol-Meyer Squibb Co. to list diabetes mellitus as a contraindication to gatifloxacin use in the US product labeling and Health Canada to issue an advisory against the use of gatifloxacin in patients with diabetes (Jose et

*Corresponding author. E-mail: nadianeuro@yahoo.com.

Author(s) agree that this article remain permanently open access under the terms of the [Creative Commons Attribution License 4.0 International License](http://creativecommons.org/licenses/by/4.0/)

al., 2007). Gatifloxacin showed to be equivalent to ciprofloxacin for the treatment of acute uncomplicated lower urinary tract infections (Naber et al., 2004). In extensive *in vivo* and *in vitro* experiments performed in an attempt to explain the central nervous system (CNS) side effects of quinolones sometimes observed under therapeutic conditions, they are like dizziness, restlessness, tremor, insomnia, hallucinations, convulsions, anxiety and depression. However, the molecular target or receptor for such effects is still not exactly known. Extensive toxicological and biochemical experiments were performed to explain the CNS effects observed under therapeutic conditions (Akahane et al., 1993; De Sarro et al., 1999, De Sarro and De sarro 2001).

Seizure activity is associated with a wide range of local biochemical changes, affecting various neurotransmitters (monoamines, amino acids) (Freitas et al., 2004; Cavaleiro et al., 2006). Cortex and hippocampus areas appeared to be important in the expression of early convulsive seizures (Kelly et al., 1999; Ang et al., 2006) in addition to the important functional association between cortical regions and the hippocampus in seizure propagation (Kelly et al., 2002) and suggested playing a role in inducing convulsions by quinolones (Motomura et al., 1991). The US Food and Drug Administration (FDA) Safety Announcement (8-15-2013) has recently issued a warning about fluoroquinolone antibacterial drugs; serious side effect of peripheral neuropathy may occur soon after these drugs are taken and may be permanent.

The study designed using single oral dose of the tested quinolones to explore its neurotoxicity as the single dose in accordance with previous studies where it was used for treatment (Loo et al., 1985), randomized controlled trials (Boy et al., 2004; Kaushik et al., 2010; Heidari Bateni et al., 2014) and its prophylactic activity (Terzi et al., 2005; Alborzi et al., 2008). The study aims to ascertain the effect of oral single dose of either Cip or Gati in male albino rat on the concentrations of amino acid and monoamine neurotransmitters and acetylcholinesterase activities in the frontal cortex and hippocampus brain areas and the histopathological examination of both areas, in addition to the determination of serum glucose level.

MATERIALS AND METHODS

Experimental animals

This study carried was out on thirty-six adult male albino rats (*Rattus norvegicus*) with average body weight of range 100 ± 20 g obtained from the Egyptian Institution of Serum and Vaccine (Helwan). The experiment was conducted in the Department of Physiology in National Organization for Drug Control and Research (NODCAR). The male albino rats were housed in iron mesh cages

with seven rats each. Clean sawdust was used to keep the animals dry and clean throughout the experimental period. The experimental animals were allowed acclimating under the laboratory conditions two weeks before the beginning of the experiments. The animals were kept under controlled temperature of 21°C and 12 h light/12 h dark cycle throughout the course of experiment. A commercial pelleted diet was used during the experiment and allowed water *ad libitum*.

Drugs

Ciprofloxacin (Cipro) ($C_{17}H_{18}FN_3O_3 \cdot HCl \cdot H_2O$), manufactured by Bayer healthcare pharmaceuticals, Ciprofloxacin hydrochloride tablets and Gatifloxacin (TEQUIN) ($C_{19}H_{22}FN_3O_4 \cdot 1.5 H_2O$) manufactured by Bristol-Myers Squibb Company. The antibiotics were administered by gastric intubation technique and doses calculated equivalent to the human therapeutic dose according to the Guidance for Industry and Reviewers (2002).

Experimental design

Animals were divided into three groups using random selection; the first group (n = 12 rats) was administered 2 ml of distilled water, the second group (Cip) was administered 80 mg/100 g body weight ciprofloxacin dissolved in 2 ml water. The Gati-treated rat groups (n = 48 rats) was administered 32 mg/100 g body weight gatifloxacin dissolved in 2 ml water. Animals were sacrificed after 12 h from dose administration by rapid decapitation. Blood samples were collected and sera separated for assessment of glucose using the BioAssay Systems' glucose assay kit (QuantiChrom™ Glucose Assay Kit). The brains were dissected out quickly, weighed and cleaned. Four brains from each treated group were served for the histopathological examination according to Bancroft et al. (1996) and the rest eight brains for the biochemical analysis. The frontal cortex and hippocampus areas were separated and divided into two halves; the first half was served for acetylcholinesterase activity assay according to the modification of Ellman et al. (1961) method as described by Gorun et al. (1978). The second half was homogenized in 75% high performance liquid chromatography (HPLC) methanol (1/10 weight/volume) using a homogenizer surrounded with an ice jacket. The homogenates were used for the determination of the brain contents of amino acids using the precolumn PTC derivatization technique according to method of Henrikson and Meredith (1984) and monoamines neurotransmitters according to method described by Pagel et al. (2000).

Statistical analysis

Reported values were represented as means \pm SE. Statistical analysis was evaluated by one-way ANOVA. Once a significant F-test was obtained, least significance difference (LSD) comparisons was performed to assess the significance of differences among various groups using statistical processor system support "SPSS" for Windows software, Release 20.0 (SPSS, Chicago, IL).

RESULTS

The data as presented in Figure 1 as percentage change from control about frontal cortex showed decrease in

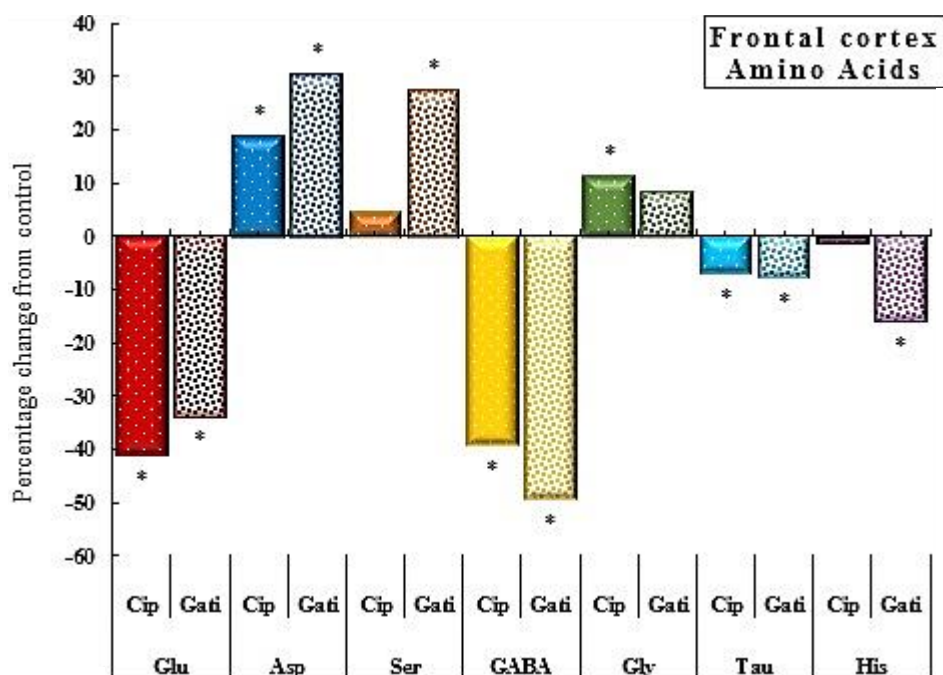


Figure 1. Percentage change from control of amino acids in cortices of rats treated with either ciprofloxacin (Cip) (80 mg/kg) or gatifloxacin (Gati) (32 mg/kg) single dose.

levels of glutamic acid, GABA, taurine and histidine and increase in aspartic, glycine and serine levels post administration of either ciprofloxacin or gatifloxacin. In ciprofloxacin group, glutamic acid, GABA, taurine and histidine data as mean \pm SE is given as 5.32 ± 0.17 (-40.97%), 1.51 ± 0.05 (-39.02%), 1.86 ± 0.04 (-6.85%) and 1.20 ± 0.01 (-1.15%), respectively, while in gatifloxacin group, results are given as 5.99 ± 0.15 (-33.60%), 1.26 ± 0.003 (-49.21%), 1.85 ± 0.06 (-7.45%) and 1.03 ± 0.02 (-15.76%) different from control values 9.02 ± 0.25 , 2.47 ± 0.04 , 2.00 ± 0.05 and 1.22 ± 0.03 , respectively. Aspartic acid increased significantly after ciprofloxacin and gatifloxacin: 3.37 ± 0.06 (18.49%) and 2.84 ± 0.06 (30.10%), respectively, from control value 2.18 ± 0.04 . Serine level increased significantly after gatifloxacin: 0.60 ± 0.02 (27.23%) and not statistically different after ciprofloxacin administration: 0.49 ± 0.01 (4.47%), respectively, from control value 2.18 ± 0.04 . Glycine level increased significantly after ciprofloxacin: 1.90 ± 0.04 (10.97%) and not statistically different after gatifloxacin administration 1.85 ± 0.06 (8.05%) from control value 1.71 ± 0.06 .

The data represented in Figure 2 as percentage change from control about hippocampus showed no statistically different decrease in glutamic acid level and significant decrease in level of aspartic acid in

ciprofloxacin group recording 9.17 ± 0.20 (-6.72%) and 1.53 ± 0.06 (-20.67%), respectively from control values 9.83 ± 0.30 , 1.93 ± 0.05 , respectively. Meanwhile, serine, GABA, glycine and histidine levels increased significantly in ciprofloxacin group recording values, 0.25 ± 0.005 (11.89%), 2.46 ± 0.08 (27.67%), 1.14 ± 0.01 (13.91%) and 2.70 ± 0.09 (9.65%), respectively from control values 0.23 ± 0.01 , 1.92 ± 0.07 , 1.00 ± 0.02 and 2.47 ± 0.07 , respectively. Amino acids level in hippocampus of gatifloxacin group showed no statistically different increase in glutamic acid recording: 10.39 ± 0.26 (5.60%). Meanwhile, it showed significant increase in all detected amino acids recording values as: 1.98 ± 0.03 (2.37%), 0.28 ± 0.01 (22.03%), 3.17 ± 0.10 (64.59%), 1.44 ± 0.05 (44.04%), 3.62 ± 0.07 (7.75%) and 2.71 ± 0.06 (10.02%) for aspartic, serine, GABA, glycine, taurine and histidine, respectively.

Monoamines level and acetylcholinesterase activities recorded percentage change from control in frontal cortex and hippocampus of treated groups as well as serum glucose presented in Figure 3. In ciprofloxacin group, noradrenaline increased significantly in frontal cortex and hippocampus as mean \pm SE by 1.19 ± 0.03 (12.44%) and 0.65 ± 0.03 (15.60%), respectively. While in gatifloxacin group, noradrenaline decreased significantly in frontal cortex recording 0.78 ± 0.03 (-26.48%) and significantly

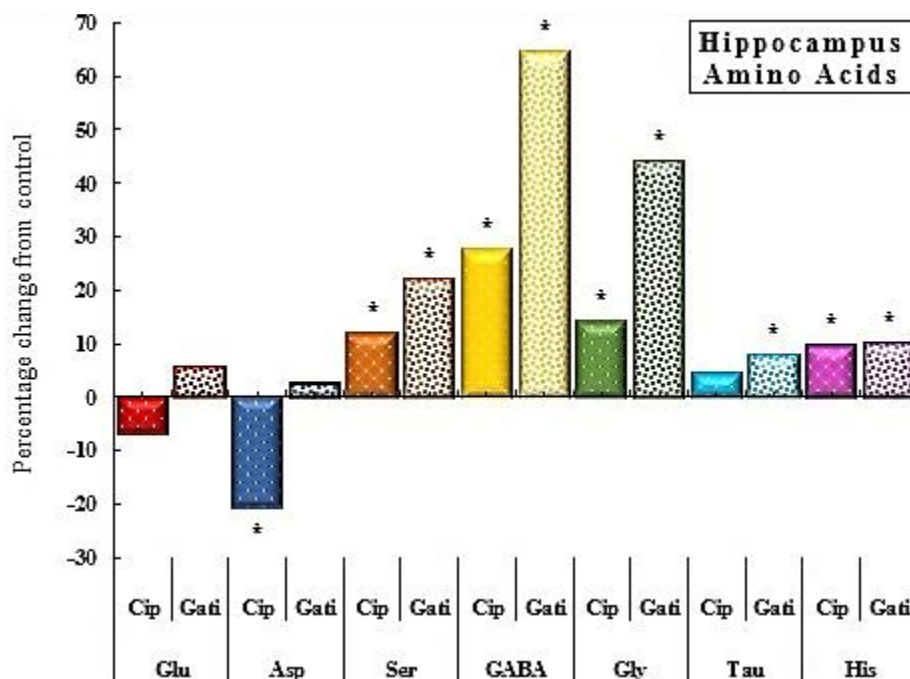


Figure 2. Percentage change from control of amino acids in hippocampi of rats treated with either ciprofloxacin (Cip) (80 mg/kg) or gatifloxacin (Gati) (32 mg/kg) single dose.

increased in hippocampus 0.87 ± 0.01 (54.79) from control values 1.06 ± 0.004 and 0.56 ± 0.05 , respectively. Dopamine decreased significantly in frontal cortex of gatifloxacin group 2.95 ± 0.05 (12.21%) from control value 3.36 ± 0.11 . Serotonin level decreased significantly in cortical area of ciprofloxacin and gatifloxacin groups 0.07 ± 0.001 (-15.30%) and 0.48 ± 0.002 (-43.53%), respectively from control value 0.08 ± 0.003 . Meanwhile it increased significantly in hippocampus of gatifloxacin group 0.29 ± 0.004 (19.95%) from control value 0.25 ± 0.006 .

Acetylcholinesterase activities increased significantly in frontal cortex while it decreased significantly in hippocampus of both treatments recording in frontal cortex activities of 16.7 ± 0.61 (21.82%) and 15.64 ± 0.91 (17.84%) in ciprofloxacin and gatifloxacin groups, respectively from control value 13.27 ± 0.33 . While in hippocampus the data recorded was 15.47 ± 0.55 (-11.70%) and 16.23 ± 0.61 (-7.36%) in ciprofloxacin and gatifloxacin groups, respectively from control value 17.52 ± 0.25 . In addition, serum glucose level increased, recording 16.7 ± 0.61 (21.82%) and 121.88 ± 3.50 (40.69%) in ciprofloxacin and gatifloxacin groups, respectively from control value 86.63 ± 0.93 . With regards to the hispopathological examination, the response of cortex and hippocampus cells to Cip and Gati administration is represented in Figure 4A to D. Figure 4A and B showed

normal histology of cerebral cortex and hippocampus in control group. There was no histopathological alteration observed in hippocampus of Cip group in Figure 4C, while in Gati group there was neuronal degeneration in the cerebral cortex (Figure 4D) associated with congestion in the blood vessels and capillaries of the hippocampus (Figure 4E).

DISCUSSION

Fluoroquinolones had structural similarities to kynurenic acid and other similar compounds which are endogenous ligands of the glutamate receptor, which might suggest an interaction of quinolones with ligand-gated glutamate receptors as well (Schmuck et al., 1998), and may explain the effect on quinolones subjected groups. The excitatory potency of fluoroquinolones is based on activation of the N-Methyl-d-aspartate (NMDA) receptor by abolishing the Mg^{2+} block in the ion channel which would prolong the opening time of the channel, thus increasing intracellular Ca^{2+} concentration in the neurons (Sen et al., 2007). The characteristics of gatifloxacin transport across blood brain barrier were investigated using primary cultured rat brain microvessel endothelial cells (rBMECs) as an *in vitro* model and study suggested that gatifloxacin transport across rBMECs involves a

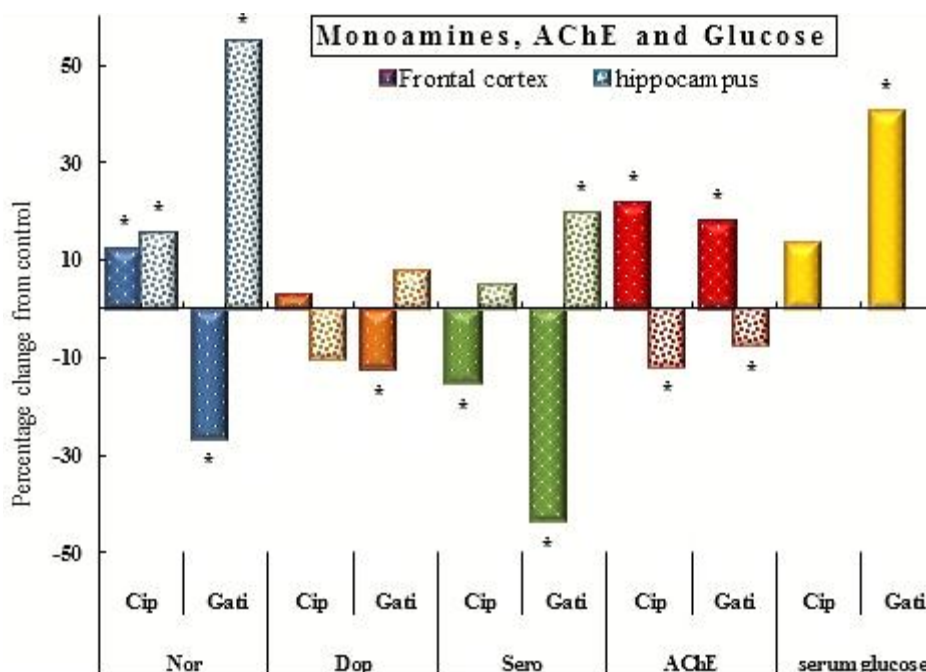


Figure 3. Percentage change from control of monoamines and acetylcholinesterase (AChE) in cortices and hippocampi and serum glucose of rats treated with either ciprofloxacin (Cip) (80 mg/kg) or gatifloxacin (Gati) (32 mg/kg) single dose.

Na⁺/Ca²⁺ exchange mechanism and extracellular Ca²⁺ (Li et al., 2009). The effect on Ca²⁺ may declare the effect of both antibiotics on taurine levels detected favoring recovery after neuronal hyperactivity (Rawi et al., 2011). Elevated aspartate, serine and glycine might suggest to the excitatory potencies of fluoroquinolones through their activation role on N-Methyl-D-aspartate-type glutamate receptor (NMDAR) (Curras and Dingledine, 1992; Wolosker, 2006; Wolosker et al., 2008).

The regional differences in GABA levels and acetylcholinesterase activities recorded decrease of GABA level and increase of AChE activity in the cortical area. Meanwhile, increase of GABA level and decrease of AChE activity in the hippocampal area in both treatments mimics that predicted in rat epileptic models (Appleyard et al., 1986) and support the proconvulsant effect of the quinolones previously discussed (Smolders et al., 2002; Abdel-Rahman et al., 2013; Arafa et al., 2013). Biochemical studies proposed role for AChE in brain mechanisms in development of status epilepticus through decrease in the AChE activity in the hippocampus (Freitas et al., 2006). The effect of ciprofloxacin and gatifloxacin on GABA levels and acetylcholinesterase activities in cortex and hippocampus and their relation to anxiety and seizure generation was discussed in our previous study (Rawi et al., 2011; Abdel-Rahman et al., 2013). Seizure induction or decrease seizure threshold

related effect to either ciprofloxacin or gatifloxacin single dose was previously declared (Darwish, 2008; Quigley and Lederman, 2004). In addition, serine elevation might be related to hippocampal serotonin increment detected in our study (Santini et al., 2014). Histidine content decreased in the frontal cortex and increased in hippocampus of ciprofloxacin and gatifloxacin treated groups. Histamine synthesis rate is a function of histidine content and histidine raises the possibility of a profound direct effect on CNS function (Yoshimatsu et al., 2002) and the herein results support the anaphylactoid reactions and hypotensive action of quinolones under therapeutic conditions as reported by Furuhashi et al. (1998), Johannes et al. (2007) and Jones et al. (2013).

Fluoroquinolone-associated anaphylaxis may occur after first-ever intake of the agent (Sachs et al., 2006). In addition, drugs that release histamine may provoke headache, asthma, hypotension, arrhythmia, urticaria, pruritus, flushing and other conditions in patients with histamine intolerance (Maintz and Novak, 2007). Monoamines levels recorded in the tested antibiotics shows elevation in noradrenaline and reduction in dopamine and serotonin in the frontal cortex in the Cip and Gati groups except reduced level of noradrenaline in Gati group. However, in hippocampus there are elevations in monoamines levels in both groups except reduction of dopamine level in Gati group. These data

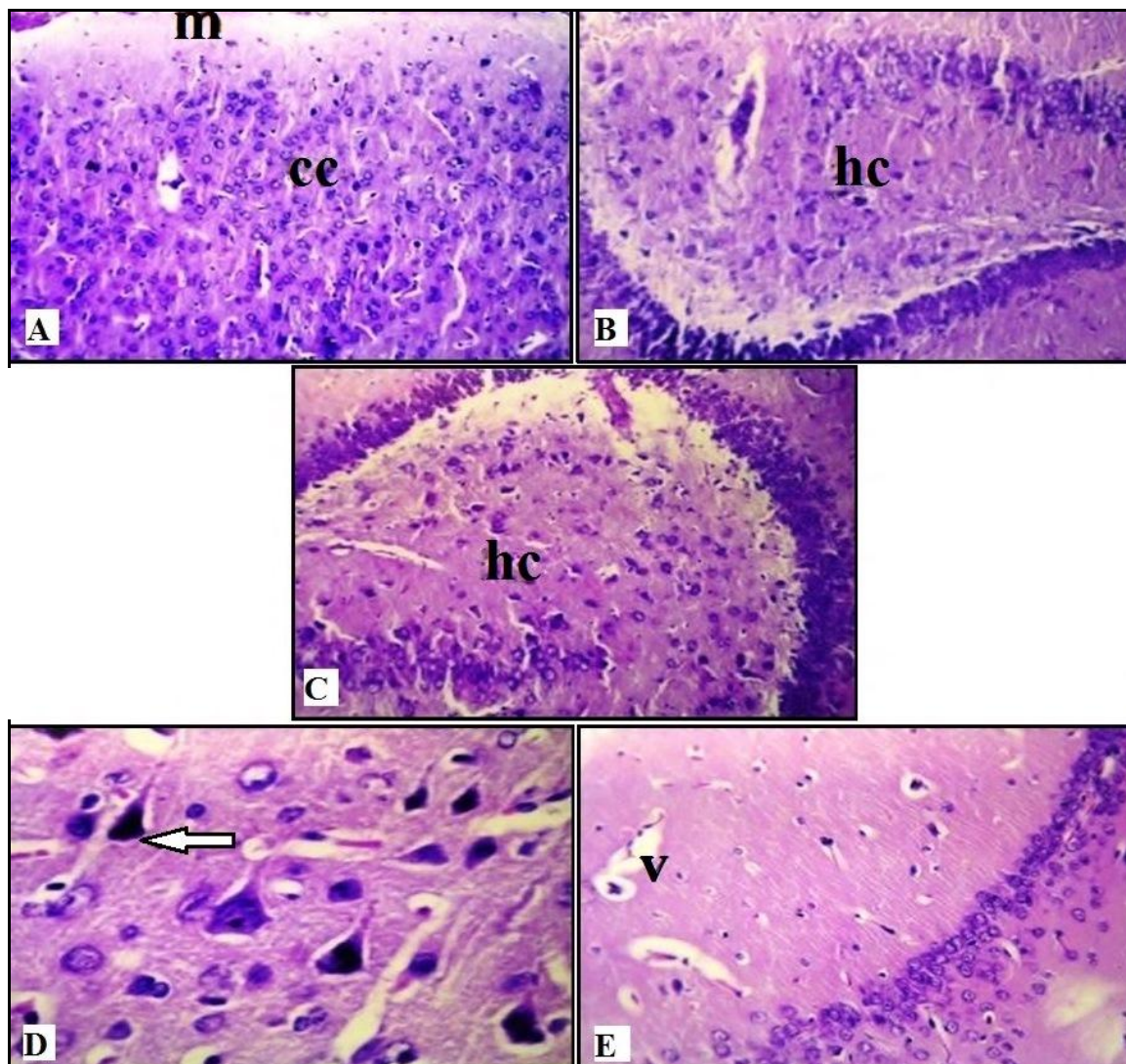


Figure 4. Light micrographs of brain sections in treated groups showing in control group normal histology of cerebral cortex (CC) and covering meninges (m) (H&E \times 40) (A) and normal rat histology of hippocampus (hc). (H&E \times 40) (B). Normal histological structure observed in the hippocampus tissue (hc) in ciprofloxacin group. (H&E \times 40) (C). Neuronal degeneration in the cerebral cortex (arrow). (H&E \times 160) (D), and congestion in the blood vessels and capillaries of the hippocampus (H&E \times 64) (E) in gatifloxacin group.

may be validated by the seizure induction through the assumption about the pharmacological treatments that lowering monoamine levels in the brain generally increase the susceptibility to seizures, while treatments that increase monoamines decrease the susceptibility (Kiyofumi and Akitane, 1977). The data recorded about monoamines in the tested antibiotics may be a supplement data to the previously mentioned seizure inducing activity of quinolones (Ooie et al., 1997; Moorthy et al., 2008; Agbaht et al., 2009). The involvement of prefrontal cortex in depression and the link between

reduced serotonin level in prefrontal cortex and depression symptoms as previously stated (Juckel et al., 1999; Koenigs and Grafman, 2009) is in accordance with the levels detected in our study. So the increment in the intracellular Ca^{2+} ions led to the rupture of the vesicles in the presynaptic terminals and increased the release of the neurotransmitters (Bullock et al., 1995) as a result, the content of catecholamine is decreased. The neurotransmitters alterations support the hyperexcitability which reflected on the histopathology of cortex and hippocampus mainly in the most affected Gati group in

line with several previous studies discussed in Rawi et al. (2011) and Arafa et al. (2013).

As regard to the effect on glucose level in tested groups, Yamada et al. (2006) reported the effect of gatifloxacin on insulin secretion and islet insulin content by using isolated mouse pancreatic islets. Islet insulin content significantly decreased by gatifloxacin already at day one; however, there are some case reports that show that only one or two doses of gatifloxacin can induce hyperglycemia (Biggs, 2003; Arce et al., 2004). Gatifloxacin was withdrawn from clinical use after reports of drug-induced hyperglycemia and other fluoroquinolones reported to interfere with glucose homeostasis (Telfer, 2014). Onyenwenyi et al. (2008) indicated that non-diabetic gatifloxacin treated patients appeared to have an increased risk of hyperglycemia and the risk reduced in diabetics. Ghaly et al. (2009) previously documented that fluoroquinolones did not stimulate insulin secretion in the presence of a basal glucose concentration; rather, they only enhanced the secretion elicited by a stimulatory glucose concentration.

Recent study by Ghaly et al. (2014) explained why fluoroquinolones produce hypo- and hyperglycaemias, because fluoroquinolones affect the function of the mitochondria in pancreatic beta cells, which may diminish the insulinotropic effect of KATP channel closure and contribute to the hyperglycaemic episodes. In addition, ciprofloxacin and gatifloxacin cause oxidative stress and decrease the mitochondrial membrane potential (Lowe et al., 2009; Talla and Veerareddy, 2011; Rawi et al., 2011; Arafa et al., 2013). Gatifloxacin acutely diminish gluconeogenesis by inhibition of mitochondrial pyruvate transport (Drozak et al., 2008) since pancreatic beta cells have an exceedingly low antioxidant capacity (Lenzen et al., 1996) and inhibition of pyruvate transport may interfere with nutrient stimulation of insulin secretion. The study of Telfer (2014) extended to suggest a connection between the ingestion of fluoroquinolones antibiotics and the development of type 2 diabetes and advice that follow-up longitudinal studies to be undertaken to examine the history of individual diabetic patients for previous fluoroquinolone exposure. Glucose resuscitation resulting in hyperglycaemia activates the NADPH pathway in neurons, causing cytotoxic oxidative stress. The same phenomenon could also adversely affect oligodendrocytes (Suh et al., 2007). The study concluded from the recorded results that the excitatory potency of ciprofloxacin and gatifloxacin could be achieved from the first dose.

ACKNOWLEDGEMENT

The authors gratefully acknowledge the Department of physiology, NODCAR, Egypt.

Conflict of interests

The authors have declared that no competing interest exists.

REFERENCES

- Abdel-Rahman M, Arafa NM, El-Khadragy MF, Kassab RB (2013). The neuroprotective role of *Nigella sativa* extract on ciprofloxacin and pentylentetrazole treated rats. *Afr. J. Pharm. Pharmacol.* 7(24):1660-1670.
- Agbaht K, Bitik B, Piskinpasa S, Bayraktar M, Topeli A (2009). Ciprofloxacin-associated seizures in a patient with underlying thyrotoxicosis: case report and literature review. *Int. J. Clin. Pharmacol. Ther.* 47(5):303-310.
- Akahane K, Kato M, Takayama S (1993). Involvement of Inhibitory and Excitatory Neurotransmitters in Levofloxacin- and Ciprofloxacin-Induced Convulsions in Mice. *Antimicrob. Agents Chemother.* 37(9):1764-1770.
- Alborzi A, Oskoe S, Pourabbas B, Alborzi S, Astaneh B, Gooya MM, Kaviani MJ (2008). Meningococcal carrier rate before and after hajj pilgrimage: effect of single dose ciprofloxacin on carriage. *East Mediterr. Health J.* 14(2):277-282.
- Ang CW, Carlson GC, Coulter DA (2006). Massive and specific dysregulation of direct cortical input to the hippocampus in temporal lobe epilepsy. *J. Neurosci.* 26(46):11850-11856.
- Appleyard ME, Green AR, Smith AD (1986). Acetylcholinesterase activity in regions of the rat brain following a convulsion. *J. Neurochem.* 46:1789-1793.
- Arafa NM, Abdel-Rahman M, El-Khadragy MF, Kassab RB (2013). Evaluation of the Possible Epileptogenic Activity of Ciprofloxacin: The Role of *Nigella sativa* on Amino Acids Neurotransmitters. *Neurochem. Res.* 8:174-185.
- Arce FCA, Bhasin RS, Pasmantier RM (2004). Severe hyperglycemia during gatifloxacin therapy in patients without diabetes. *Endocr. Pract.* 10:40-44.
- Banchroft JD, Stevens A, Turner DR (1996). Theory and practice of histological techniques. Fourth Ed. Churchill Livingstone, New York, London, San Francisco, Tokyo. Elsevier Health Sciences p 125.
- Biggs WS (2003). Hypoglycemia and hyperglycemia associated with gatifloxacin use in elderly patients. *J. Am. Board Fam. Pract.* 16:455-457.
- Boy D, Well M, Kinzig-Schippers M, Sörgel F, Ankel-Fuchs D, Naber KG (2004). Urinary bactericidal activity, urinary excretion and plasma concentrations of gatifloxacin (400 mg) versus ciprofloxacin (500 mg) in healthy volunteers after a single oral dose. *Int. J. Antimicrob. Agents* 23(1):6-16.
- Bullock J, Boyle J, Wang MB (1995). Synaptic transmission. In: *Physiology Middle East Edition* 3rd ed, Chapter 3, Williams & Wilkins. London pp. 22-31.
- Cavalheiro EA, Fernandes MJ, Turski L, Naffah-Mazzacoratti MG (2006). Spontaneous recurrent seizures in rats: Amino acid and monoamine determination in the hippocampus. *Epilepsia.* 35(1):1-11.
- Curras MC, Dingle R (1992). Selectivity of amino acid transmitters acting at N-methyl-D-aspartate and amino-3-hydroxy-5-methyl-4-isoxazolepropionate receptors. *Mol. Pharmacol.* 41:520-526.
- Darwish T (2008). Ciprofloxacin-induced seizures in a healthy patient. *N. Z. Med. J.* 121(1277):104-105.
- De Sarro A, De sarro G (2001). Adverse reactions to fluoroquinolones. An overview on mechanistic aspects. *Cur. Med. Chem.* 8(4):371-384.
- De Sarro A, Cecchetti V, Fravolini V, Naccari F, Tabarrini O, De sarro G (1999). Effects of novel 6-desfluoroquinolones and classic quinolones on pentylentetrazole-induced seizures in mice. *Antimicrob. Agents. Chemother.* 43(7):1729-1736.
- Drozak J, Miecznik A, Jarzyna R, Bryla J (2008). The inhibition of gluconeogenesis by gatifloxacin may contribute to its hypoglycaemic action

- Eur. J. Pharmacol. 594:39-43.
- Ellman GL, Courtney KD, Andres V, Featherstone RM (1961). A new and rapid colorimetric determination of cholinesterase activity. *Biochem. Pharmacol.* 7:88-95.
- Freitas RM, Sousa FC, Viana GS, Fonteles MM (2006). Acetylcholinesterase activities in hippocampus, frontal cortex and striatum of Wistar rats after pilocarpine-induced status epilepticus. *Neurosci. Lett.* 399 (1-2):76-78. PMID: 16481111
- Freitas RM, Vasconcelos SMM, Souza FCF, Viana GSB, Fonteles MMF (2004). Monoamine levels after Pilocarpine-induced status epilepticus in hippocampus and frontal cortex of wistar rats. *Neurosci. Lett.* 370 (2-3):196-200.
- Furuhata K, Hayakawa H, Soumi K, Arai H, Watanabe Y, Narita H (1998). Histamine-releasing properties of T-3762, a novel fluoroquinolone antimicrobial agent in intravenous use. I. Effects of doses and infusion rate on blood pressure, heart rate and plasma histamine concentration. *Biol. Pharm. Bull.* 21(5):456-460.
- Ghaly H, Jörns A, Rustenbeck I (2014). Effect of fluoroquinolones on mitochondrial function in pancreatic beta cells. *Eur. J. Pharm. Sci.* 52:206-214.
- Ghaly H, Kriete C, Sahin S, Pflöger A, Holzgrabe U, Zünkler BJ, Rustenbeck I (2009). The insulinotropic effect of fluoroquinolones. *Biochem. Pharmacol.* 77:1040-1052.
- Gorun V, Proinov I, Baltescu V, Balaban G, Barzu O (1978). Modified Ellman procedure for assay of cholinesterase in crude enzymatic preparation. *Anal. Biochem.* 86:324-326.
- Guidance for Industry and Reviewers (2002). Estimating the safe starting dose in clinical trials for therapeutics in adult healthy volunteers. Food and Drug Administration 1-26. <http://www.fda.gov/OHRMS/DOCKETS/98fr/02d-0492-gdl0001-vol1.pdf>
- Heidari Bateni Z, Shahrokh H, Salimi H, Safari H, Tabatabai M, Saedi D (2014). Single-Dose versus Multiple-Dose Ciprofloxacin plus Metronidazole Prophylaxis in Transrectal Ultrasound-Guided Biopsy of the Prostate: a Randomized Controlled Trial. *Acta Med. Iran.* 52(9):664-670.
- Heinrikson RL, Meredith SC (1984). Amino acid analysis by RP-HPLC: precolumn Derivatization with phenylisothiocyanate. *Anal. Biochem.* 136:65-74.
- Johannes CB, Ziyadeh N, Seeger JD, Tucker E, Reiter C, Faich G (2007). Incidence of Allergic Reactions Associated with Antibacterial Use in a Large, Managed Care Organisation. *Drug Saf.* 30(8):705-713.
- Jones SC, Budnitz DS, Sorbello A, Mehta H (2013). US-based emergency department visits for fluoroquinolone-associated hypersensitivity reactions. *Pharmacoepidemiol. Drug Saf.* 22(10):1099-1106.
- Jose J, Jimmy B, Saravu K (2007). Dysglycemia associated with the use of fluoroquinolones-focus on gatifloxacin. *J. Clin. Diagn. Res.* (3):185-187.
- Juckel G, Mendlin A, Jacobs BL (1999). Electrical stimulation of rat medial prefrontal cortex enhances forebrain serotonin output: implications for electroconvulsive therapy and transcranial magnetic stimulation in depression. *Neuropsychopharmacology* 21(3):391-398.
- Kaushik JS, Gupta P, Faridi MM, Das S (2010). Single dose azithromycin versus ciprofloxacin for cholera in children: a randomized controlled trial. *Indian Pediatr.* 47(4):309-315.
- Kelly ME, Batty RA, McIntyre DC (1999). Cortical spreading depression reversibly disrupts convulsive motor seizure expression in amygdala-kindled rats. *Neuroscience* 91:305-313.
- Kelly ME, Staines WA, McIntyre DC (2002). Secondary generalization of hippocampal kindled seizures in rats: examination of the periform cortex. *Brain Res.* 957(1):152-161.
- Kiyofumi K, Akitane M (1977). Brain Monoamines in Seizure Mechanism (Review). *Folia Psychiatr. Neurol. Jpn.* 31(3):483-489.
- Koenigs M, Grafman J (2009). The functional neuroanatomy of depression: distinct roles for ventromedial and dorsolateral prefrontal cortex. *Behav. Brain Res.* 201(2):239-243.
- Lenzen S, Drinkgern J, Tiedge M (1996). Low antioxidant enzyme gene expression in pancreatic islets compared with various other mouse tissues. *Free Radic Biol. Med.* 20(3):463-466.
- Li Y, Liu L, Li J, Xie L, Wang GJ, Liu XD (2009). Transport of gatifloxacin involves Na⁺/Ca²⁺ exchange and excludes P-glycoprotein and multidrug resistance associated-proteins in primary cultured rat brain endothelial cells. *Eur. J. Pharmacol.* 616(1-3):68-72
- Loo PS, Ridgway GL, Oriel JD (1985). Single dose ciprofloxacin for treating gonococcal infections in men. *Genitourin Med.* 61(5):302-305.
- Lowes DA, Wallace C, Murphy MP, Webster NR, Galley HF (2009). The mitochondria targeted antioxidant MitoQ protects against fluoroquinolone-induced oxidative stress and mitochondrial membrane damage in human Achilles tendon cells. *Free Radic Res.* 43(4): 323-328.
- Maintz L, Novak N. (2007). Histamine and histamine intolerance. *Am. J. Clin. Nutr.* 85(5):1185-1196.
- Moorthy N, Raghavendra N, Venkatarathnamma PN (2008). Levofloxacin-induced acute psychosis. *Indian J. Psychiatry* 50(1):57-58.
- Motomura M, Kataoka Y, Takeo G, Shibayama K, Ohishi K, Nakamura T, Niwa M, Tsujihata M, Nagataki S (1991). Hippocampus and frontal cortex are the potential mediatory sites for convulsions induced by new quinolones and non-steroidal anti-inflammatory drugs. *Int. J. Clin. Pharmacol. Ther. Toxicol.* 29(6):223-227.
- Naber KG, Allin DM, Clarysse L, Haworth DA, James IG, Raini C, Schneider H, Wall A, Weitz P, Hopkins G, Ankel-Fuchs D (2004). Gatifloxacin 400 mg as a single shot or 200 mg once daily for 3 days is as effective as ciprofloxacin 250 mg twice daily for the treatment of patients with uncomplicated urinary tract infections. *Int. J. Antimicrob. Agents* 23(6):596-605.
- Onyenwenyi AJ, Winterstein AG, Hatton RC (2008). An evaluation of the effects of gatifloxacin on glucose homeostasis. *Pharm. World Sci.* 30(5):544-549.
- Ooie T, Terasaki T, Suzuki H, Sugiyama Y (1997). Quantitative brain microdialysis study on the mechanism of quinolones distribution in the central nervous system. *Drug Metab. Dispos.* 25(7):784-789.
- Pagel P, Blome J, Wolf HU (2000). High-performance liquid chromatographic separation and measurement of various biogenic compounds possibly involved in the pathomechanism of Parkinson's disease. *J. Chromatogr. B Biomed. Sci. Appl.* 746(2): 297-304.
- Quigley CA, Lederman JR (2004). Possible gatifloxacin-induced seizure. *Ann. Pharmacother.* 38(2):235-237.
- Rawi SM, Arafat NMS, El-Hazmi MM (2011). Evaluation of the Effects of Ciprofloxacin or Gatifloxacin on Neurotransmitters Levels in Rat Cortex and Hippocampus. *Afr. J. Pharm. Pharmacol.* 5:993-1005.
- Sachs B, Riegel S, Seebeck J, Beier R, Schichler D, Barger A, Merk HF, Erdmann S (2006). Fluoroquinolone-associated anaphylaxis in spontaneous adverse drug reaction reports in Germany: differences in reporting rates between individual fluoroquinolones and occurrence after first-ever use. *Drug Saf.* 29(11):1087-1100.
- Santini MA, Balu DT, Puhl MD, Hill-Smith TE, Berg AR, Lucki I, Mikkelsen JD, Coyle JT (2014). D-serine deficiency attenuates the behavioral and cellular effects induced by the hallucinogenic 5-HT_{2A} receptor agonist DOI. *Behav. Brain Res.* 259:242-246.
- Schmuck G, Schürmann A, Schlüter G. (1998). Determination of the excitatory potencies of fluoroquinolones in the central nervous system by an *in vitro* model. *Antimicrob. Agents Chemother.* 42(7):1831-1836.
- Sen S, Jaiswal AK, Yanpallewar S, Acharya SB (2007). Anxiogenic potential of ciprofloxacin and norfloxacin in rats. *Singapore Med. J.* 48(11):1028-1032.
- Smolders I, Gousseau C, Marchand S, Couet W, Ebinger G, Michotte Y (2002). Convulsant and Subconvulsant Doses of Norfloxacin in the Presence and Absence of Biphenylacetic Acid Alter Extracellular Hippocampal Glutamate but Not Gamma-Aminobutyric Acid Levels in Conscious Rats. *Antimicrob. Agents Chemother.* 46(2):471-477.
- Suh W S, Gun ET, Hamby AM, Chan PH, Swanson RA (2007). Hypo-

- glycemic neuronal death is triggered by glucose reperfusion and activation of neuronal NADPH oxidase. *J. Clin. Invest.* 117(4):910–918.
- Talla V, Veerareddy P (2011). Oxidative stress induced by fluoroquinolones on treatment for complicated urinary tract infections in Indian patients. *J. Young Pharm.* 3(4):304-309.
- Telfer SJ (2014) Fluoroquinolone antibiotics and type 2 diabetes mellitus. *Med. Hypotheses.* 83(3):263-269.
- Terzi C, Kiliç D, Unek T, Hoşgörler F, Füzün M, Ergör G (2005). Single-dose oral ciprofloxacin compared with single-dose intravenous cefazolin for prophylaxis in inguinal hernia repair: a controlled randomized clinical study. *J. Hosp. Infect.* 60(4):340-347.
- Wolosker H, Dumin E, Balan L, Foltyn VN (2008). D-amino acids in the brain: D-serine in neurotransmission and neurodegeneration. *FEBS J.* 275(14):3514-3526.
- Wolosker H (2006). D-serine regulation of NMDA receptor activity. *Sci STKE* (356):pe41.
- Yamada C, Nagashima K, Takahashi A, Ueno H, Kawasaki Y, Yamada Y, Seino Y, Inagaki N (2006). Gatifloxacin acutely stimulates insulin secretion and chronically suppresses insulin biosynthesis. *Eur. J. Pharmacol.* 553(1-3):67-72.
- Yoshimatsu H, Chiba S, Tajima D, Akeh Y, Sakata T (2002). Histidine suppresses food intake through its conversion into neuronal histamine. *Exp. Biol. Med.* (Maywood). 227(1):63-68.

Full Length Research Paper

Drug use pattern in out-patient children: A comparison between primary and secondary health care facilities in Northern Nigeria

Basheer A. Z. Chedi^{1*}, Ibrahim Abdu-Aguye² and Helen O. Kwanashie²

¹Department of Pharmacology, Bayero University, Kano, Nigeria.

²Department of Pharmacology and Therapeutics, Ahmadu Bello University, Zaria, Nigeria.

Received 30 December, 2014; Accepted 23 January, 2015

Children are more vulnerable to adverse events related to use of drugs. It is therefore important to study drug use in children in order to optimize pharmacotherapy. The aim of this study was to compare drug utilization in paediatric outpatient departments of primary and secondary health care facilities. The patient and drug information of 600 patients was analyzed for World Health Organization (WHO) recommended prescribing indicators. The average number of drugs per prescription was significantly ($p < 0.0005$) lower in secondary (2.97) compared to primary (3.62) facilities, while average consultation time was shorter ($p < 0.0005$) in primary than secondary facilities. Percentages of drugs prescribed from Nigerian Essential Drug List (EDL, primary {89.78%}; secondary {91.79%}) and by generic name (primary {55.04%}; secondary {57.88%}) were insignificantly different between the facilities. The use of injectables was low (8.32% in primary versus 3.74% in secondary facilities) while antibiotic use was high (54.14% in primary to 60.28% in secondary facilities). Analysis of the dispensing indicators showed that the secondary facilities were significantly ($p < 0.05$) better than the primary facilities, even though not a single drug was adequately labeled in both the primary and secondary facilities. Prescription from EDL was found to be fair in the study area while use of injections was low. There is a need for improvement in case of medicines prescribed by generic name.

Key words: Drug utilization, out-patients, children, health care facility.

INTRODUCTION

Rational drug use has been defined as using the right drug in the right patient, for the right indication, in the right dose and dosage form, for the right duration of time. The rational use of drugs seeks to avoid the frequent problems of over- and under-prescription, inappropriate prescription and the use of new, expensive drugs when equally effective, well tried, safe, high quality and

cheaper alternatives are available (NEDP, 1993). Unfortunately, in most cases, prescribing and dispensing patterns do not always conform to these criteria. The consequences of such inappropriate use of drugs cannot be overlooked especially in children. This is because children differ greatly from adults, not merely in size but also in the proportions and constituents of their bodies as

*Corresponding author. E-mail: b2chedi@yahoo.com.

Author(s) agree that this article remain permanently open access under the terms of the [Creative Commons Attribution License 4.0 International License](http://creativecommons.org/licenses/by/4.0/)

Table 1. Characteristic of the general outpatient department prescribers.

Facility type	Total	Doctors [n (MYT)]	Nurses [n (MYT)]	CHO/CHEW [n (MYT)]
Secondary (n=8)	38	18 (5.2)	14 (20.4)	6 (13.3)
Primary (n=12)	39	0 (0)	0 (0)	39 (13.4)
Total [n (%)]	77	18 (23.4)	14 (18.2)	45 (58.4)

CHO: Community health officer; CHEW: community health extension worker; MYT: mean years of training.

well as functioning of their physiological systems. These differences are reflected in the way the body handles and responds to drugs and are relevant to medication (Laurence et al., 1999). No single rule or formula suffices for all paediatric cases; the dose is therefore established partly by scaling for body weight and/or surface area and by making pharmacokinetic and pharmacodynamic measurements when opportunities arise.

Studies conducted in most developing countries have shown that a high percentage of consultations end with prescriptions regardless of the necessity to prescribe (Kroenke, 1985). Numerous studies have also described irrational patterns of drug use that include polypharmacy, use of drugs that were not related to the diagnosis, patient non-compliance, overuse and misuse of antibiotics, and unnecessary use of injectable drugs (Quick et al., 2002). Such practices may result in a waste of resources, inappropriate patient demand, antimicrobial resistance, and increased drug-related morbidity and mortality. This study is therefore aimed at investigating the prescribing and dispensing practices at a representative sample of health care facilities in Kano State, Nigeria using WHO drug use indicators and comparing the results obtained between the primary and secondary health care facilities in the state.

METHODOLOGY

Study design

This was a comparative, cross sectional study involving paediatric outpatient departments of twenty (8 secondary and 12 primary) public health care facilities selected by multistage sampling technique in Kano State Northwestern Nigeria. The study was conducted between June 2009 and December 2009. The Ethical Committee of the Aminu Kano Teaching Hospital and Kano State Ministry of Health approved the study protocol.

Inclusion and exclusion criteria

The patients included in this study were those presented with general illness, aged 11 years and below. Patients presenting to the health care facilities for follow-up of chronic diseases, and patients presenting to receive services such as vaccination, and other specialized care services, were excluded.

Study sample

Based on the WHO recommended methodology (WHO, 1993),

stratification of health care facilities according to senatorial districts and systematic random sampling were used to select a total of 20 health care facilities across the state. In each health care facility, 30 paediatric outpatient prescriptions were collected using systematic random sampling.

Data collection

The principal investigator (PI) collected data on both prescribing and dispensing indicators prospectively from each facility due to difficulties encountered in the availability of retrospective records. Demographic data of each patient, diagnosis, antimicrobial sensitivity test and detailed prescription were recorded on a modified WHO core prescription indicators form. Data collected were coded and de-identified. These recorded forms were used to analyze the average number of drugs per prescription, number of encounters with antibiotics, percentage of drugs prescribed by generic name/listed in the Nigerian Essential Drug List (EDL), percentage of drugs actually dispensed and adequately labeled. The consultation and dispensing times (WHO, 1993) were collected using a disguise technique (the investigator stayed outside the consultation room and the pharmacy). At each facility, the consultation and dispensing times were counted for 30 patients, and 30 parents were interviewed upon exit after the drug(s) had been dispensed to investigate dispensing practices and patient knowledge.

Analysis

Data were entered into Microsoft excel 2007 and Statistical Package for Social Sciences (SPSS version 15) and a descriptive analysis was performed. Drug utilization indicators were computed and compared between the primary and secondary facilities by unpaired Student's t-test. $P < 0.05$ was considered to be statistically significant.

RESULTS

In total, 77 health care providers prescribed at paediatric outpatients of the 20 selected facilities during the study period out of which only 23.4% (18/77) were qualified medical doctors (Tables 1 and 2). Among the 600 patients that participated in the study, 44.7% (286) were male and the average age of the patients was 3.7 ± 0.3 years. A total of 2016 drugs were prescribed in 600 prescriptions, giving an average of 3.36; and the range of drugs per encounter varied from 2 to 5. Three drugs were prescribed in most of the patients (57.3%) and there was not a single prescription wherein no drug was prescribed. Values of the prescription indicators are as shown in Figures 1 to 6. Dispensing and other patient care indi-

Table 2. Characteristic of the general outpatient department dispensers.

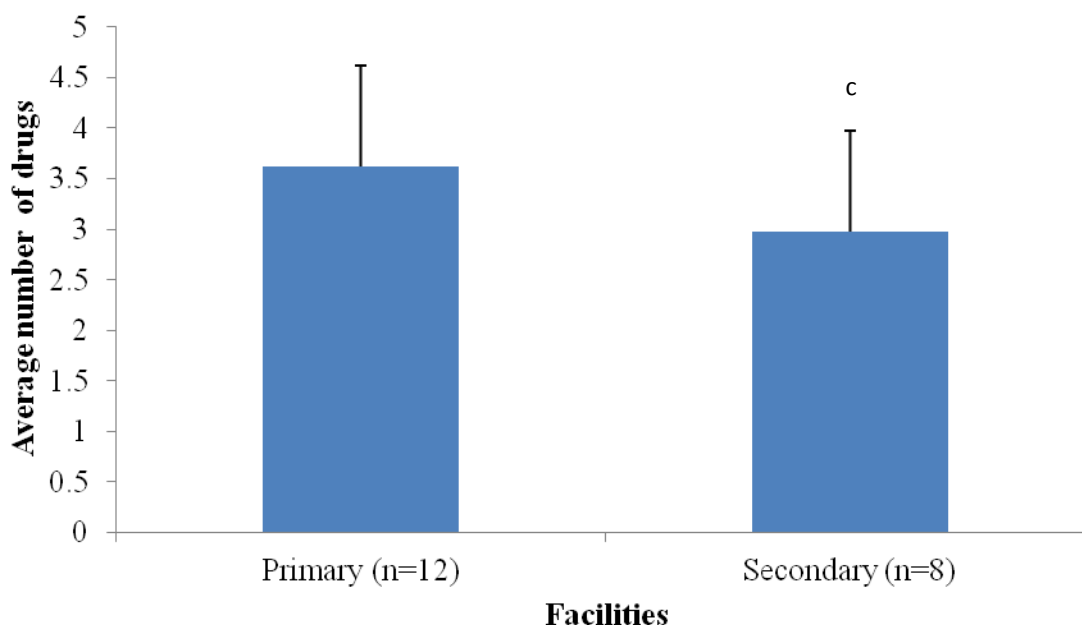
Facility type	Total	Pharmacists [n (MYT)]	Pharm Tech [n (MYT)]	CHO/CHEW [n (MYT)]
Secondary (n=8)	51	6 (6.8)	42 (13.2)	3 (7.3)
Primary (n=12)	5	0 (0)	0 (0)	5 (12.8)
Total [n (%)]	56	6 (10.7)	42 (75.0)	8 (14.3)

Pharm Tech: Pharmacy technician; CHO: community health officer; CHEW: community health extension worker; MYT: mean years of training.

Table 3. Dispensing indicators for paediatric outpatients' departments of 20 health care facilities in Kano State, Nigeria.

Indicator	Health care facilities	
	Primary (n=12)	Secondary (n=8)
Average dispensing time (s)	25.73 ± 1.18	38.68 ± 2.17 ^b
Percentage of drugs actually dispensed	56.17 ± 2.36	95.91 ± 2.08 ^c
Percentage of drugs adequately labeled	0.00 ± 0.00	0.00 ± 0.00 ^{ns}
Percentage of parents who claim to have had adequate knowledge	79.03 ± 1.45	89.26 ± 2.08 ^a

ns: Not significant; a ≤ 0.05; b ≤ 0.005; c ≤ 0.0005.

**Figure 1.** Average number of drugs per prescription for paediatric outpatients' departments of 20 health care facilities in Kano State, Nigeria. ns: Not significant; c = < 0.0005.

cators for the primary and secondary facilities are listed in [Table 3](#).

DISCUSSION

This study revealed that the mean number of drugs prescribed per patient was significantly lower ($p < 0.0005$) in secondary (2.97) compared to primary facilities (3.62).

Values of 2.3 to 3.7 drugs per encounter in secondary facilities have been reported from India (Dimri et al., 2008), Nigeria (Odusanya, 2004) and Ghana (Owusu-Dakuu and Sablah, 2004). In primary facilities, rates of 2.5 to 3.13 drugs per encounter have been reported in India (Anuja et al., 2010), Nigeria (Nwolisa et al., 2006) and Yemen (Bashrahil, 2010). In the present study, three or more drugs were prescribed in 72.4% of prescriptions, which reflects a trend towards polypharmacy, as it has

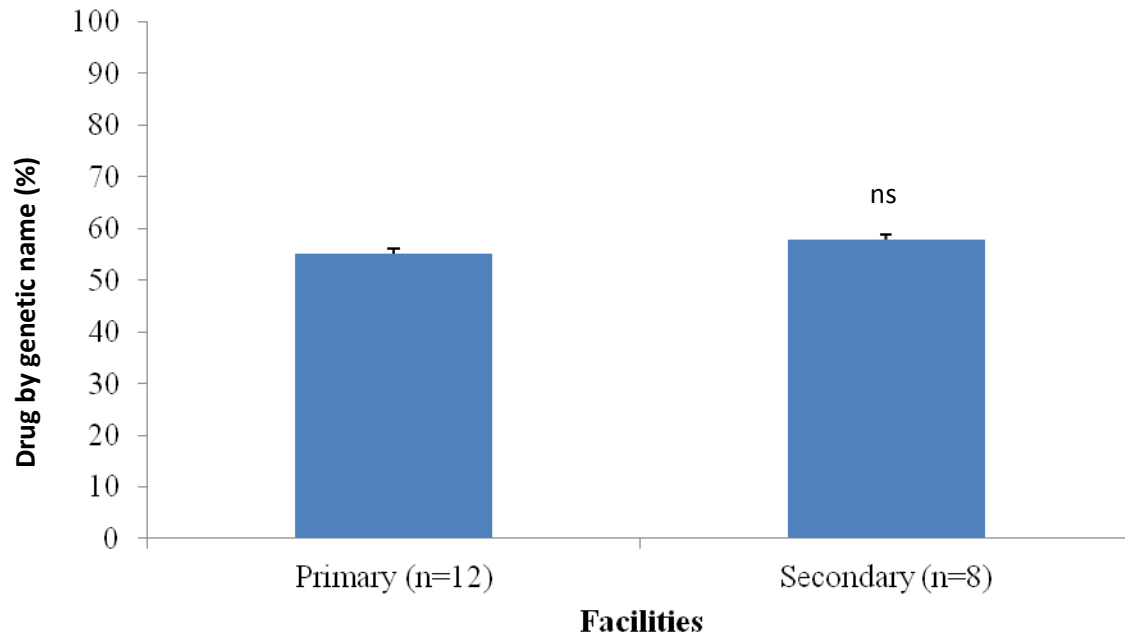


Figure 2. Percentage of drugs prescribed by generic name for paediatric outpatients' departments of 20 health care facilities in Kano State, Nigeria. ns: Not significant.

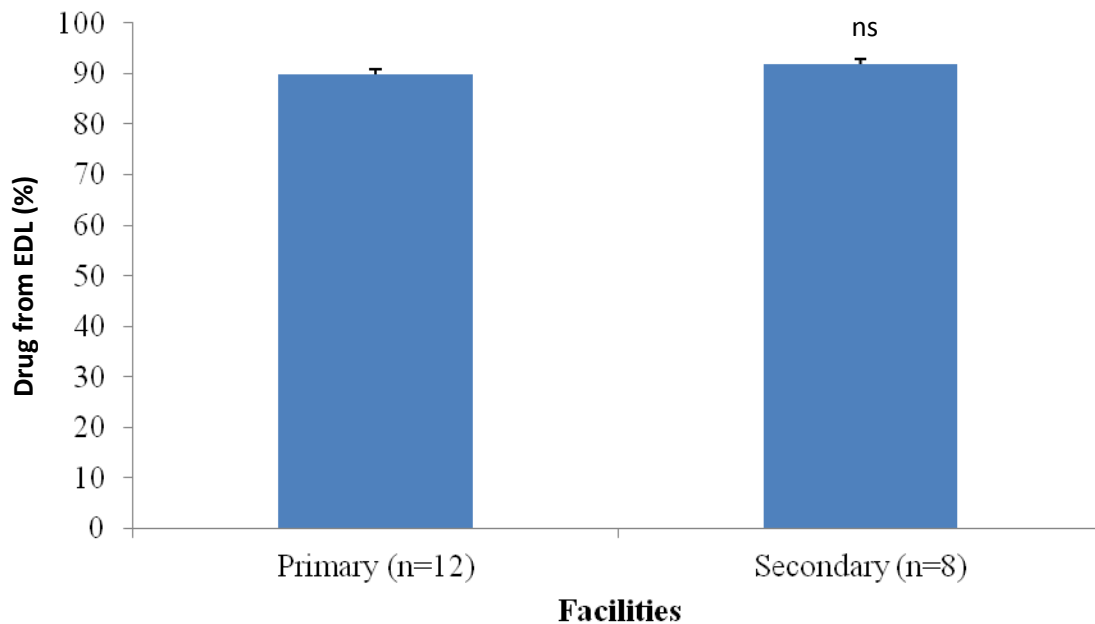


Figure 3. Percentage of drugs prescribed from EDL for paediatric outpatients' departments of 20 health care facilities in Kano State, Nigeria. ns: Not significant.

been proposed that the average number of drugs per prescription should be 1.6 to 1.8 (Isah et al., 2006). Prescriptions of high numbers of drugs may be attributed to patient demand; patients believe that the prescribing of more drugs will ensure improvement and facilitate the

cure of their conditions more quickly. Other possible factors could be that the treatment is based on pure symptoms instead of a proper diagnosis due to lack of adequately trained personnel, laboratory facilities and overcrowdings in the health care centres. Although

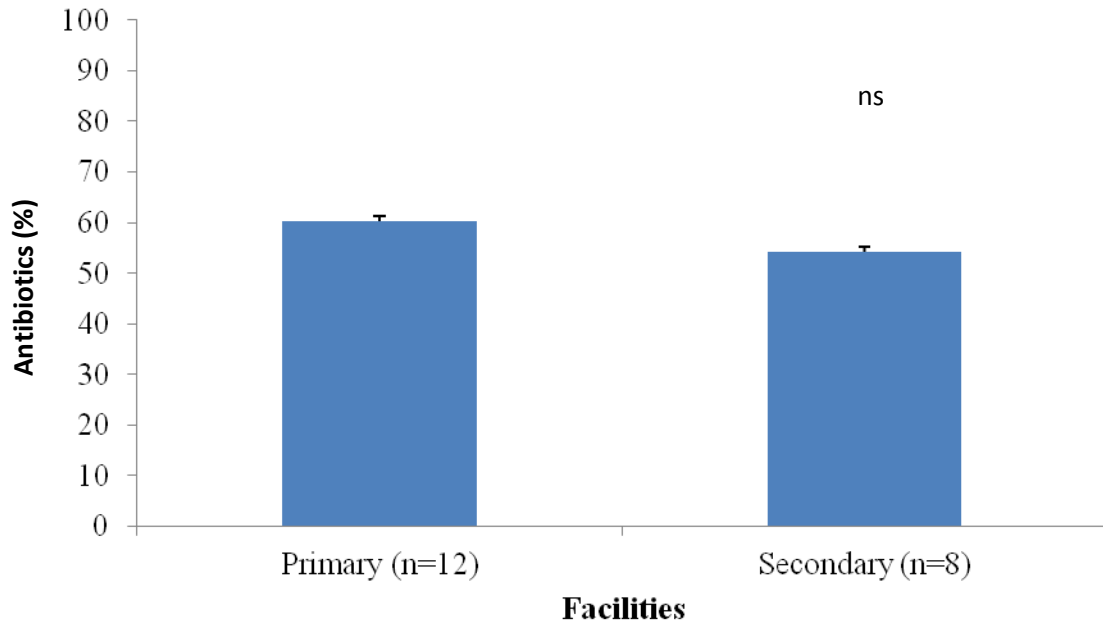


Figure 4. Percentage of antibiotics prescribed per encounter for paediatric outpatients' departments of 20 health care facilities in Kano State, Nigeria. ns: Not significant.

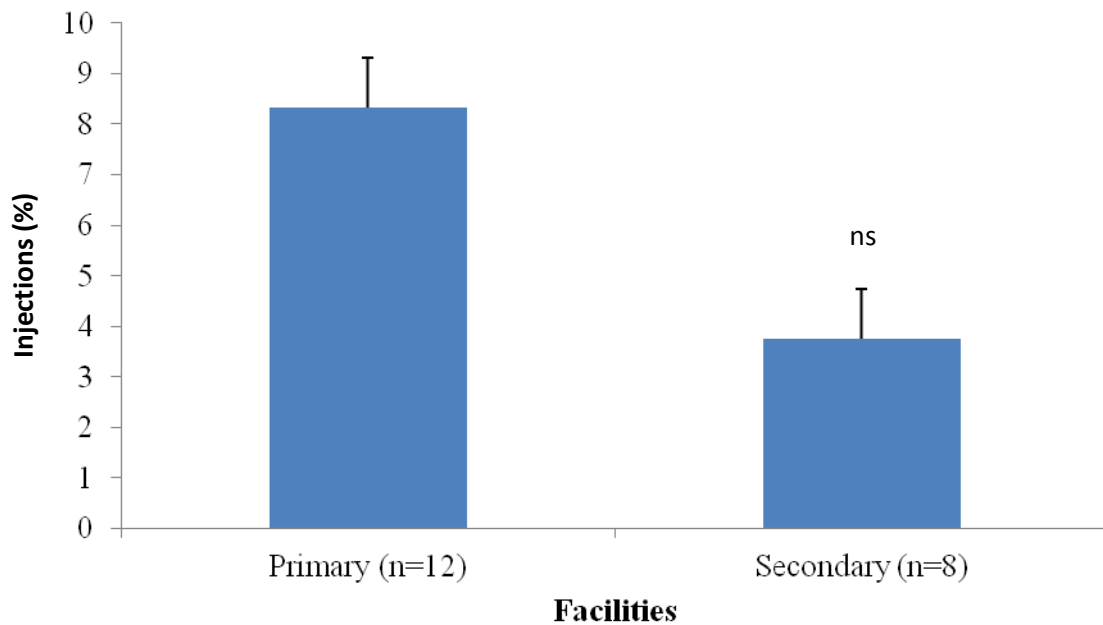


Figure 5. Percentage of injections prescribed per encounter for paediatric outpatients' departments of 20 health care facilities in Kano State, Nigeria. ns: Not significant.

increasing efforts are being made to improve drug-use practices in developing countries, it should be noted that some of these patients visit the hospitals with other diseases such as anaemia, malnutrition and at times some other infections. These make poly pharmacy inevitable.

Prescribing by generic name is known to reduce the

cost of drug treatment and rationalizing drug therapy. This varies from 13.3 to 93% across the globe (Nsimba, 2006). Prescribing by generic name in our study was similar (55 to 61%) in most facilities studied. Other studies have reported even lower percentages, ranging from 25 to 60%, (Bashrahil, 2010; Chedi et al., 2010), while the optimal percentage should be close to 100%

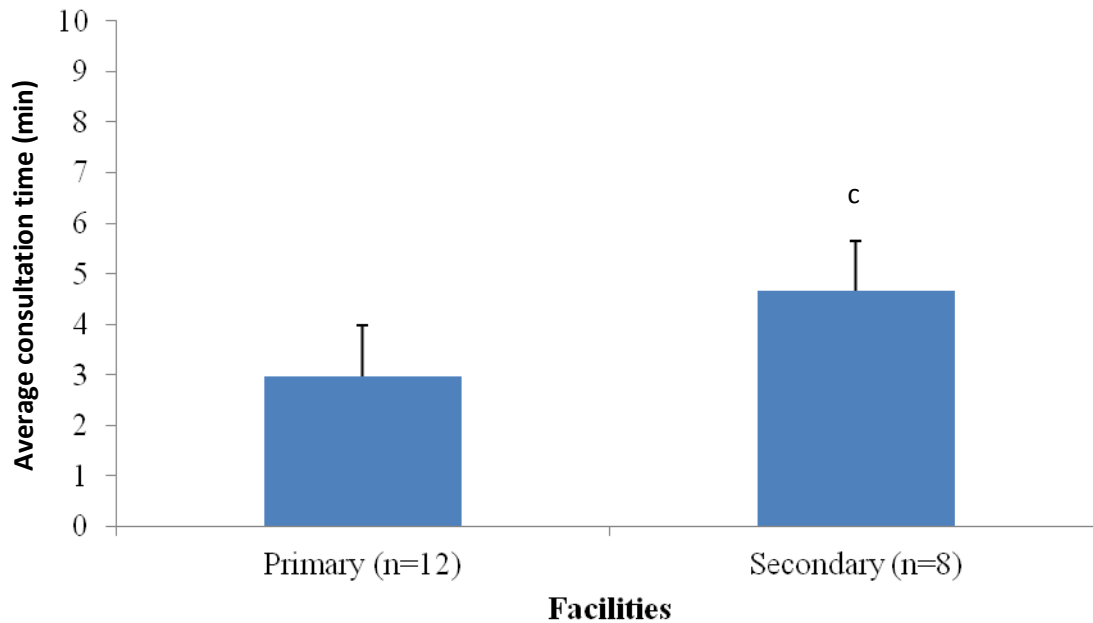


Figure 6. Average consultation time (min) for paediatric outpatients' departments of 20 health care facilities in Kano State, Nigeria. ns: Not significant; $c \leq 0.0005$.

(Isah et al., 2006). The low rate of generic prescribing may be due to the fact that the information on medicines that prescribers usually receive is mostly provided by the drug companies. Another contributing factor for low generic prescribing by prescribers is their beliefs that the brand names are easy to remember as well as the frequent use of brand names during their training (Awad et al., 1999). Generic prescribing is strongly recommended, as it facilitates education and knowledge, and it also allows the pharmacist to maintain a more economic stock control system based on a smaller range of reasonably priced drugs (Awad and Himad, 2006).

The percentages of drugs prescribed from EDL in this study were lower among the primary facilities (89.78%) compared with secondary facilities (91.79%) but this was not significant ($p > 0.05$). These figures were higher than the values previously reported in the same area (Chedi et al., 2010) but lower than the optimal value of 100% (Isah et al., 2006). When the prescribers were asked concerning their low number of prescriptions from the EDL, some of the arguments forwarded by them were that, not all the drugs for various diseases are available in the EDL and resistance had developed to some of the drugs on the list. Appropriate use of antibiotics is necessary to prevent emergence of drug resistant bacteria. It has been recommended that fewer than 30% of prescriptions should contain an antibiotic (Isah et al., 2006). Percentages of antibiotics prescribed per encounter obtained in this study compared favourably with those reported from Ghana (Bosu and Ofori-Adjei, 2000), Cambodia (Chareonkul et al., 2002) and other parts of Nigeria (Chukwuani et al., 2002). The major

factors that influence high antibiotic prescribing at health centers have been reported to be a lack of knowledge about appropriate antibiotic use, including overestimation of the severity of illness to justify antibiotic prescribing by prescribers, and pressure from patients who believe that antibiotics provide rapid symptomatic relief of the disease (Awad et al., 2006). The same factors may possibly play a role in health care centres of Kano. Antibiotics are essential drugs, but the overuse may increase antibiotic resistance, which will endanger their therapeutic effectiveness, increase treatment failure and, as a result, lead to longer and more severe illness episodes with higher costs and mortality rates.

Although inappropriate high levels of injection prescribing (17.1% to 80%) have been reported in Tanzania (Massele et al., 2001) and Zimbabwe (Trap et al., 2002), in the present study, overall injection use was low (5.0 to 8.0%). The proportion was higher in primary as compared to secondary facilities, but not significant. During the period of this study, prescribers spent a mean of 4.3 to 5.0 min in the secondary and 2.9 to 3.1 min in the primary facilities with patients. These figures correspond well with values measured in other developing countries (Hogerzeil et al., 1993). Health care providers have attributed the short time of contact with patients to their work overload, namely, a large number of patients. Though it is difficult to present optimal standards for consultation time, prescribers should take enough time to provide the patient with the necessary information regarding his/her condition and instructions/warnings related to the prescribed drug.

The mean dispensing time was found to be shorter ($p <$

0.005) in primary (24.8 to 36.1 s) than that observed in secondary (35.8 to 44.8 s) facilities. These figures are slightly higher than the average value obtained (12.5 s) from other studies in twelve developing countries (Hogerzeil et al., 1993) but far shorter than 86.1 s recorded in Nepalese pharmacies (Kafle et al., 1992). Although, from a management point of view, a short dispensing time could imply a very efficient dispensing system, from a clinical point of view, such a short time would not be expected to provide adequate counseling on medication, and may infer less attention to detail and a greater potential for errors. Factors that may account for a short dispensing time include pre-packed and pre-labeled drugs as well as a heavy flow of patients or even shortage of staff.

The percentage of drugs that were actually dispensed in secondary facilities was close to 100%, as recommended (Isah et al., 2006). This indicates a good stock control system in the secondary health care centers in Kano. In contrast, only 56.17% was actually dispensed in the primary facilities studied. The significant difference ($p < 0.0005$) between the secondary and primary facilities could be explained by the fact that the Drug Revolving Fund (DRF) scheme of all the secondary health care facilities in the state was supported by the Partnership for Transformation in the Health Sector in Nigeria (PATH), while only one primary facility out of the twelve selected was under the programme during the study period. Although the WHO (1993) and National Drug Policy of Nigeria (NDP) (2005) recommend that each drug label should contain the dose regimen, generic name of the drug and patient's name. In the study area, not a single dispensed drug was adequately labeled in all the facilities. Similar result was obtained in Cambodia (Chareonkul et al., 2002) and India (Rishi et al., 2003). At the end of the study, when the dispensers were asked about the inadequate labeling, they stated that given their typical workload they hardly had time to interact with the parents; hence, they prefer to draw pictograms and explain how the individual drugs should be taken.

About 90 and 80% of parents in the secondary and primary health care facilities, respectively claimed to know the correct dosage schedule. These figures, though higher than 55 to 68.3% reported in Bangladesh (Guyon et al., 1994), Burkina Faso (Krause et al., 1999), Cambodia (Chareonkul et al., 2002) and India (Rishi et al., 2003), did not necessarily reflect reality since the response "Yes, I know the dose" was accepted as positive answer.

Conclusion

Conclusively, this study provides few insights into the drug use patterns in children outpatient departments of primary and secondary public health care facilities in Kano State, Nigeria. The prescription from EDL was fair, the use of injections was low and there is a scope for

improvement in case of medicines prescribed by generic name.

Conflict of interest

Authors declare that there are no conflicts of interest

REFERENCES

- Anuja AP, Subhash BT, Prakash RB (2010). Prescription Analysis of Pediatric Outpatient Practice in Nagpur City. *Indian J. Comm. Med.* 35(1):70-73.
- Awad AI, Himad HA (2006). Drug use practices in teaching hospitals of Khartoum State, Sudan. *Eur. J. Clin. Pharmacol.* 62:1087-1093
- Awad AI, Eltayeb IB, Baraka OZ (2006). Changing antibiotics prescribing practices in health centers of Khartoum State, Sudan. *Eur. J. Clin. Pharmacol.* 3:1-8
- Awad AI, El-Tayeb IB, Omer ZB (1999). Investigation of drug use in health centers in Khartoum State. *Sudan Med. J.* 37:21-26.
- Bashrahil KA (2010). Indicators of rational drug use and health services of anti-malarial drugs utilization in public health facilities in Kano, Nigeria. *Bayero J. Pure Appl. Sci.* 3(1):49-53
- Chukwuani CM, Onifade M, Sumonu K (2002). Survey of drug use practices and antibiotic prescribing pattern at a general hospital in Nigeria. *Pharm. World Sci.* 24:188-195.
- Dimri S, Tiwari P, Basu S, Parmar VR (2008). Drug Use Pattern in Children at a Teaching Hospital. *Indian Paediatr.* 46:165-167
- Guyon AB, Barman A, Ahmed JU, Ahmed AU, Alam MS (1994). A baseline survey on use of drugs at the primary health care level in Bangladesh. *Bull. World Health Organ.* 72: 265-271.
- Hogerzeil HV, Bimo, Ross-Degnan D, Laing RO, Oforie-Adjei D, Santoso B, Azad Chowdury AK, Das AM, Kafle KK, Mabadaje AFB (1993). Field test for rational drug use in twelve developing countries. *Lancet* 342:1408-10.
- Isah AO, Ross-Degnan D, Quick J, Laing R, Mabadaje AFB (2009). The development of standard values for the WHO drug use prescribing indicators. International Conference on Improving Use of Medicines, 2006. Available at: http://archives.who.int/icium/icium1997/posters/1a2_txt.html
- Kafle KK, Karkee SB, Prasad RR (1992). INRUD drug use indicators in Nepal: practice patterns in health post in four districts. *INRUD News* 3:15.
- Krause G, Borchert M, Benzler J (1999). Rationality of drug prescriptions in rural health centers in Burkina Faso. *Health Policy Plan* 14:291-298.
- Kroenke K (1985). Polypharmacy. Causes, consequences and cure. *Am. J. Med.* 79:149-152.
- Laurence DR, Bennet PN, Brown MJ (1999). General Pharmacology. In: Laurence DR, Bennet PN and Brown MJ (Ed) *Clinical Pharmacology*, 8th edn. Churchill and Livingstone, Edinburgh London p 112.
- Massele AY, Nsimba SE, Rimoy G (2001). Prescribing habits in church-owned primary health care facilities in Dar Es Salaam and other Tanzanian coast regions. *East Afr. Med. J.* 78:510-514.
- NDP (2005). National Drug Policy. National Formulary and Essential Drugs Review Committee, FMOH Document, Abuja, Nigeria.
- NEDP (1993). National Essential Drug Policy. National Formulary and Essential Drugs Review Committee, FMOH Document, Abuja, Nigeria.
- Nsimba SE (2006). Assessing prescribing and patient care indicators for

- children under five years old with malaria and other disease conditions in public primary health care facilities. *Southeast Asian J. Trop. Med. Public Health* 37:206-214.
- Nwolisa CE, Erinaugha EU, Ofoleta SI (2006). Prescribing practices of doctors attending to under-fives in a children's outpatient clinic in Owerri, Nigeria. *J. Trop. Pediatr.* 52(3):197-200.
- Odusanya OO (2004). Drug use indicators at a secondary health care facility in Lagos, Nigeria. *J. Comm. Med. Primary Health Care* 16(1):21-24.
- Owusu-Dakuu FTK, Sablah J (2004). The essential drug list and drug use indicators at two university hospitals: KNUST and Legon, Ghana. *West Afr. J. Pharm.* 18(1):53-57.
- Quick JD, Hogerzeil HV, Velasquez G, Rago L (2002). Twenty-five years of essentials medicines. *Bull World Health Organ* 80:913-914.
- Rishi RK, Sangeeta S, Surendra K, Tailang M (2003). Prescription audit: experience in Garhwal (Uttaranchal), India. *Trop. Doct.* 33:76-79.
- Trap B, Hansen EH, Hogerzeil HV (2002). Prescription habits of dispensing and non-dispensing doctors in Zimbabwe. *Health Policy Plan* 17:288-295.
- World Health Organization and International Network for Rational Use of Drugs (1993). How to investigate drug use in health facilities: Selected drug use indicators. EDM Research Series No. 7 [WHO/DAP/93.1]. Geneva: World Health Organization, 1993; IOS Press

Full Length Research Paper

Performance, parasitic infections, hematology and hepatic histology of *Colossoma macropomum* (tambaqui) fed on homeopathic product

Douglas Anadias Pinheiro¹, Bruno Adan Sagratzki Caverio¹, Lauro Vargas², Graciela Lucca Braccini², Eliane Tie Oba Yoshioka³, Marcos Sidney Brito Oliveira³ and Marcos Tavares-Dias^{3*}

¹Universidade Federal do Amazonas, Programa de Pós-Graduação em Ciências Pesqueiras nos Trópicos, Manaus, AM, Brazil.

²Universidade Estadual de Maringá, Departamento de Zootecnia, Maringá, PR, Brazil.

³Embrapa Amapá, Laboratório de Sanidade de Organismos Aquáticos, Macapá, AP, Brazil.

Received 1 October, 2014; Accepted 30 January, 2015

Homeopathic products may act in an organism by stimulating the immune system, allowing the restoration of balance and encouraging organic response under stress. This study investigated the performance, blood, morphological and parasitological parameters in *Colossoma macropomum* (tambaqui) fed diets containing different concentrations of Homeopatila 100[®]. Juvenile tambaqui underwent four treatments with three replicates: 0, 20, 40 and 60 ml Homeopatila 100[®]/kg of extruded feed with 32% crude protein for 60 days. At the end of 60 days the growth performance, blood parameters, gill parasites and hepatic histology in fish fed homeopathic product were evaluated. Treatment with Homeopatila 100[®] did not improve the growth performance of fish. There was no difference in the prevalence and abundance of monogeneans and protozoans in the gills of fish, except in those fed with 60 ml homeopathic product. The plasma glucose levels were higher in fish fed diet containing 40 and 60 ml homeopathic product. The mean corpuscular volume and hematocrit levels in fish fed 20 and 60 ml were higher than in controls. In fish with fed 40 ml increased number of neutrophils and reduced number of lymphocytes was found. However, 60 ml of the product caused increase in the number of monocytes and reduced the number of lymphocytes, eosinophils and PAS-positive granular leukocytes. Under conditions in this study, the Homeopatila 100[®] did not improve fish performance or reduce parasitic infections, but showed a relative improvement in blood response of fish fed on 40 ml of this homeopathic complex.

Key words: Fish farming, homeopathy, monogenea, parasites, protozoa, blood.

INTRODUCTION

Currently, almost half of the fish production comes from aquaculture in Brazil, but due to demand for products

*Corresponding author. E-mail: marcos.tavares@embrapa.br.

Author(s) agree that this article remain permanently open access under the terms of the [Creative Commons Attribution License 4.0 International License](http://creativecommons.org/licenses/by/4.0/)

based on fish, the production will increase in the coming decades, mainly by socioeconomic and health reasons. The country has a huge potential for expansion of this activity, such as favorable climate, possibility of use of Union's water for fish farming, both reservoirs and estuary, plus a large number of biodiversity species with livestock potential (Rocha et al., 2013). Moreover, it produces native species of great economic interest as *Colossoma macropomum* (tambaqui), a Serrasalminidae from the Amazon River basin.

Tambaqui is the most cultivated native fish in Brazil; its production was 111,084.1 tons in 2011, accounting for 20.4% of total domestic production. This production represents an increase of over 100% compared to 2010 (MPA, 2013). This increase in domestic production is mainly due to its zootechnical characteristics that favor the production of this Amazonian fish, such as rapid growth, relative resistance to diseases and good tolerance to high temperature and to low levels of dissolved oxygen in the water (Araujo-Lima and Gomes, 2005; Santos et al., 2013). It has regular supply of fingerlings and good yield of fillet without skin, its flesh is of good nutritional quality and its production cycle in cages is short, six to eight months (Oliveira et al., 2013).

Growth in fish farming is one of the most important and commonly used criteria to measure the fish response to the diet or ingredients used in the feed. Because growth is the measure of greatest applicability in the production systems to assess the growth performance of cultured fish, once it is closely related to productivity and profitability (Fracalossi et al., 2013), being assessed in different ways. With the intensification of the tambaqui fish farming, the disease problems have increased mainly by the infections of protozoan *Ichthyophthirius multifiliis* and monogenean helminthes, which may impair the production and productivity (Tavares-Dias et al., 2013). Such problems of parasitic diseases require constant need of treatment to reduce and control the parasites during intensive production of this fish. Prophylactic care should be permanent in fish farming of tambaqui, due to the difficulty of treating infectious and parasitic diseases when installed. Homeopathic products may act in the body of animals by stimulating the immune system, allowing the restoration of balance and encouraging organic responses in reducing stress. The use of such products, besides contributing to the prophylaxis by reducing the management stress, can reduce the use of chemotherapy and antibiotics, avoiding risks to the environment, animals and consumers (Siena et al., 2010). However, the use of homeopathic products and their potential benefits are virtually unknown in fish farming.

In *Oreochromis niloticus* (Nile tilapia), 40 ml Homeopatila 100[®]/kg diet improved the survival of fingerlings, feed conversion, hepatosomatic index, increased the number of muscle fibers, number of hepatocytes and hepatic glycogen (Vargas and Ribeiro,

2009; Braccini et al., 2013). However, there are few studies on the effects of homeopathic products in fish. This study evaluated the growth performance, blood and morphological parameters and gill infections in *C. macropomum* fed diets containing different concentrations of Homeopatila 100[®].

MATERIALS AND METHODS

Experimental design

In this experiment, 300 fingerlings of *C. macropomum* (12.0 ± 0.5 cm and 42.7 ± 3.1 g) obtained from commercial fish farming (Macapá, AP, Brazil) were acclimated for seven days in water tanks. Fish were randomly distributed in water tanks (500 L), containing 400 L useful volume, maintained at a density of 25 fish per tank. The design was completely randomized with four treatments (20 ml hydroalcoholic solution - controls, 20, 40 and 60 ml Homeopatila 100[®]/kg diet) and three replications. The homeopathic product was added to the extruded commercial diet containing 32% crude protein and fish fed for 60 days.

Preparation of diets with homeopathic product

We used commercial diet containing 32% crude protein, 65 g ether extract, 70 g crude fiber, 100 g mineral matter, 12 g calcium, 6000 mg phosphorus, 16000 IU vitamin A, 250 IU vitamin E, 4500 IU vitamin D3, 30 mg vitamin K3, 325 mg vitamin C and 32 mg thymine (B1) for each kg of diet. Weekly, Homeopatila 100[®] in the form of hydroalcoholic solution was incorporated to the commercial feed using a hand sprayer. Composition of the complex Homeopatila 100[®], for 1000 ml: 250 ml of iodum 12 cH, 250 ml of sulphur 30 cH, 250 ml of natrummuriaticum 200 cH, 250 ml of streptococcinum 30 cH and q.s medium (ethylalcohol 30%). Subsequently, the feed was homogenized and dried at room temperature, removing it periodically for 24 h. The feed was stored in a cool dry place without any incidence of sunlight, chemicals and equipment that emitted magnetic field until being loose and without alcohol odor (Siena et al., 2010). The same inclusion process was conducted to the control treatment using 20 ml of 30% alcohol per kg feed. The amount of feed provided to fish was *ad libitum*, and three times a day (9:00, 13:00 and 17:00 pm).

Parameters of growth performance

In the initial and final experiment, all fish were weighed (g) in a digital scale and measured in total length (cm) using caliper to determine the following parameters of the body growth performance:

1. Weight gain (g) = $WG = (W_1 - W_2)$
2. Daily weight gain (DWG) = WG/t
3. Specific growth rate (%) = $(SGR) = 100 \times (\ln W_1 - \ln W_2) / t$
4. Feed conversion rate (FCR) = Amount / weight gain

Where W_1 = mean weight (g) in the final experiment; W_2 = mean weight (g) in the initial experiment; t = time (days) of experiment

5. Relative condition factor (Kn) (Le-Cren, 1951).

Procedures for collection and analysis of blood parameters

After 60 days of feeding with 20 ml of hydroalcoholic solution

Table 1. Physical and chemical parameters of the farming water of *Colossoma macropomum* fed on different concentrations of Homeopatila 100®.

Treatments (mg/kg)	Oxygen (mg/L)	pH	Temperature (°C)	Conductivity (µs/cm)	Total ammonia (mg/L)
Control	5.57±0.58 ^a	6.05±0.41 ^a	29.82±0.41 ^a	0.036±0.003 ^a	0.85±0.57 ^a
20	5.43±0.64 ^a	6.12±0.38 ^a	29.90±0.42 ^a	0.037±0.003 ^a	0.66±0.37 ^a
40	5.49±0.61 ^a	6.09±0.39 ^a	29.93±0.42 ^a	0.038±0.005 ^a	0.71±0.54 ^a
60	5.46±0.63 ^a	6.09±0.38 ^a	29.98±0.43 ^a	0.036±0.003 ^a	0.69±0.37 ^a

Means followed by different letters in the same column indicate differences between treatments by the Tukey test ($p < 0.05$). Values expressed as mean \pm standard deviation.

(control), 20, 40 and 60 ml Homeopatila100®/per kg feed, five fish per replicate of the different treatments were anesthetized with eugenol (15 mg/L water) (Inoue et al., 2011) for blood collection. An aliquot of blood collected by puncture of the caudal vessel was collected from the 60 fish with the aid of syringes containing Ethylenediaminetetraacetic acid (10%). The blood was used to determine the total number of erythrocytes in a Neubauer chamber, concentration of hemoglobin using Drabkin's reagent and reading on a spectrophotometer at 540 nm absorbance and hematocrit by the microhematocrit method. With this data, the Wintrobe erythrocytic indices were calculated: mean corpuscular volume (MCV) and mean corpuscular hemoglobin concentration (MCHC). Blood smears were confectioned and stained with a combination for differential leukocyte counts in up to 100 cells of interest, in each extension and also for determining the total number of leukocytes and total thrombocytes (Ranzan-Paiva et al., 2013). Leukocytes were identified and classified in lymphocytes, monocytes, neutrophils, eosinophils and PAS-positive granular leukocytes (LG-PAS), following the recommendations of Tavares-Dias et al. (1999). A portion of blood was centrifuged to obtain the plasma and determination of the concentrations of glucose and total protein using kits from Doles (GO, Brazil) and spectrophotometric reading.

Parasitological analysis

After 60 days of feeding on 20 ml hydroalcoholic solution (control), 20, 40 and 60 ml Homeopatila100® per kg diet, 10 fish of each repetition of the different treatments were anesthetized with eugenol (15 mg/water) (Inoue et al., 2011), then the gills were collected for parasitological analysis. The gills of 120 fish were collected, fixed in 5% formalin and used for collection, fixation, quantification and preparation for the identification of parasites (Eiras et al., 2006). The prevalence and mean abundance were calculated (Bush et al., 1997) for each treatment.

Biometric and histological analyses of the liver

After 60 days of feeding on 20 ml of hydroalcoholic solution (control), 20, 40 and 60 ml Homeopatila100® per kg diet, 10 fish of each replication of the different treatments were anesthetized with 15 mg/L of eugenol (Inoue et al., 2011) for liver collection. The liver of 120 fish was removed, weighed on a precision scale used to determine the Hepatosomatic index (HSI) (Tavares-Dias et al., 2000). For each treatment, liver of two other fish of each repetition was collected, totaling 24 fish, washed in sodium chloride (0.85%) and fixed in Bouin solution for 12 h. Then, the fragments were washed in 70% ethanol for removal of picric acid, dehydrated and embedded in paraffin, following routine procedures. After making the slides (in duplicate) for morphological analysis and determination of the number of hepatocytes per area, the staining

was performed with hematoxylin and eosin (Behmer et al., 1976) and for histochemical staining of hepatic glycogen (Beçak and Paulete, 1976); in order to quantify the percentage occupied by this inclusion/area, the periodic acid-Schiff (PAS) + hematoxylin was used. The morphometric analyses of liver tissue were performed from images captured on Olympus BX41 optical microscope coupled with camera Olympus Q-Color 3, using a 40x magnification objective. Fifty images of each fish were analyzed, totaling 300 images/treatment; the standardized floor area was 20914.7 µm². Such measurements were performed with the aid of image analysis of software Image Pro Plus® (Version 4.5, Media Cybernetics, USA).

Physical and chemical parameters of water in the tanks

In the experimental tanks, water renovation was constant and the accumulated debris was siphoned every two days. The water temperature, pH, dissolved oxygen levels, ammonia and electrical conductivity were measured daily, using digital devices (YSI, USA) for each purpose.

Statistical analyses

All data have been previously evaluated under assumptions of normality and homoscedasticity using Shapiro-Wilk and Bartlett tests, respectively. For data with normal distribution the analysis of variance (ANOVA - one way) was used, followed by Tukey test for means comparison. For data that did not follow this pattern of distribution the Kruskal-Wallis test was used, followed by Dunn's test for means comparison.

RESULTS

The physical and chemical parameters of the water in the farming tanks of tambaqui fed on different concentrations of Homeopatila 100® showed no differences between them (Table 1). Initial and final values of length and weight of tambaqui are in Figure 1. There was no significant difference ($p > 0.05$) among treatments after 60 days of feeding on homeopathic product. After 60 days of fish feeding, the weight gain, feed conversion ratio and specific growth rate were different only in fish kept at 60 ml Homeopatila 100®/kg diet compared to control and other treatments. However, the Kn of fish was not influenced by treatments with homeopathic product (Table 2). The gills of the fish were parasitized by

Table 2. Parameters of production performance of *Colossoma macropomum* fed on different concentrations of Homeopatila 100® for 60 days.

Treatments (ml/kg)	WG (g)	DWG(g/day)	FCR	SGR (%)	Kn	S (%)
Control	154.4 ± 34.3 ^{ab}	2.6 ± 0.57 ^a	1.52	2.51 ± 0.33 ^{ab}	0.99 ± 0.01 ^a	100
20	167.1 ± 41.3 ^{ab}	2.8 ± 0.68 ^a	1.53	2.63 ± 0.34 ^a	1.00 ± 0.01 ^a	100
40	169.8 ± 47.69 ^b	2.8 ± 0.79 ^a	1.57	2.65 ± 0.40 ^a	1.00 ± 0.03 ^a	100
60	150.57 ± 42.03 ^a	2.5 ± 0.70 ^b	1.72	2.47 ± 0.35 ^b	0.99 ± 0.01 ^a	100

WG: Weight gain; DWG: Daily weight gain; FCR: Feed conversion rate; SGR: specific growth rate; Kn: relative condition factor; S: Survival. Means followed by different letters in the different rows indicate differences between treatments by the Tukey test ($p < 0.05$). Values expressed as mean ± standard deviation.

Table 3. Prevalence (P%) and mean abundance (MA) of parasites in the gills of *Colossoma macropomum* fed different concentrations of Homeopatila 100®/kg feed for 60 days.

Parasite species	Control		20 ml/kg		40 ml/kg		60 ml/kg	
	P (%)	MA	P (%)	MA	P (%)	MA	P (%)	MA
<i>Ichthyophthirius multifiliis</i>	100	7.144.3 ± 4033.6 ^a	100	13.310.9 ± 4997.0 ^b	100	10.401.8 ± 6853.3 ^{ab}	100	54.108.3 ± 70851.9 ^c
<i>Piscinoodinium pillulare</i>	0	0 ^a	0	0 ^a	0	0 ^a	30	362.5 ± 615.43 ^b
<i>Anacanthorus spatulathus</i>	100	24.1 ± 18.0 ^a	100	40.6 ± 59.5 ^{ab}	100	24.9 ± 14.2 ^a	100	53.2 ± 33.1 ^b
<i>Notozothecium janauachensis</i>	100	27.1 ± 15.3 ^a	100	30.5 ± 30.60 ^a	100	36.0 ± 26.2 ^{ab}	100	51.9 ± 31.0 ^b
<i>Mymarothecium boegeri</i>	100	15.3 ± 4.8 ^a	100	14.7 ± 10.0 ^a	100	22.8 ± 21.0 ^a	100	43.4 ± 27.7 ^b
<i>Linguadactyloides brinkimanni</i>	23.3	0.96 ± 2.8 ^a	13.3	0.63 ± 2.3 ^a	3.3	0.1 ± 0.5 ^a	10	54.108.3 ± 70851.9 ^c

Values expressed as mean ± standard deviation. Means followed by different letters in the same row indicate difference between treatments by the Dunn test ($p < 0.05$), for mean abundance.

Ichthyophthirius multifiliis Fouquet, 1876 (Ciliophora), *Piscinoodinium pillulare* (Schäperclaus, 1954) Lom 1981 (Dinoflagellida), *Anacanthorus spatulathus* Kritsky, Thatcher and Kayton, 1979, *Notozothecium janauachensis* Belmont-Jegu, 2004, *Mymarothecium boegeri* Cohen and Kohn, 2005 and *Linguadactyloides brinkimanni* Thatcher and Kritsky, 1983 (Dactylogyridae). There was no difference in the prevalence of these parasites among groups of fish fed on different concentrations of homeopathic product. However, *P. pillulare* occurred only in fish maintained on diets containing 60 ml of Homeopatila 100® (Table 3).

In hosts, the highest parasite abundance was of *I. multifiliis* and the lowest abundance was of *L. brinkimanni*. The abundance of *I. multifiliis* was higher in fish fed on diets containing 20 and 60 ml Homeopatila 100® compared to control fish. The abundance of *A. spatulathus*, *N. janauachensis* and *M. boegeri* was higher in fish fed diets containing 60 ml of homeopathic product when compared with controls (Table 3). Plasma glucose levels were higher in fish fed on diets containing 40 and 60 ml Homeopatila 100® when compared to controls. Hematocrit and Mean corpuscular volume (MCV) of fish fed on diets containing 20 and 60 ml of homeopathic product were higher

than in controls. In fish fed diet containing 20 ml of homeopathic product, the total number of thrombocytes and monocytes were higher than in controls. However, the number of lymphocytes was reduced in all groups fed on different concentrations of homeopathic products. The number of neutrophils was increased only in fish with 40 ml Homeopatila 100® when compared to controls, while the number of PAS-LG and eosinophils were smaller in fish fed on this homeopathic product 100® (Table 4).

The percentage of glycogen did not differ among treatments with homeopathic product. However, the number of hepatocytes was lower in

Table 4. Blood parameters of *Colossoma macropomum* fed on different concentrations of Homeopatila 100®/kg feed for 60 days.

Parameters	Control	20 mL/kg	40 ml/kg	60 ml/kg
Glucose (mg dl ⁻¹)	93.4 ± 5.4 ^a	99.9 ± 8.6 ^a	110.9 ± 11.5 ^b	113.6 ± 12.4 ^b
Protein (mg dl ⁻¹)	3.6 ± 0.5 ^a	3.6 ± 0.3 ^a	3.6 ± 0.4 ^a	3.6 ± 0.4 ^a
Hematocrit (%)	19.7 ± 1.5 ^a	22.5 ± 3.7 ^b	20.7 ± 2.3 ^{ab}	22.9 ± 1.8 ^b
Hemoglobin (g/dl ⁻¹)	7.2 ± 1.3 ^a	7.25 ± 0.97 ^a	7.0 ± 0.7 ^a	7.5 ± 0.9 ^a
RBC (number 10 ⁶ /µl ⁻¹)	0.99 ± 0.17 ^a	0.87 ± 0.20 ^a	0.95 ± 0.2 ^a	0.96 ± 0.26 ^a
MCV (fL ⁻¹)	204.5 ± 33.9 ^a	263.9 ± 55.6 ^b	224.5 ± 39.3 ^{ab}	251.3 ± 55.8 ^b
MCHC (g/dl ⁻¹)	36.8 ± 6.1 ^a	32.8 ± 4.7 ^a	33.9 ± 3.6 ^a	32.8 ± 2.3 ^a
Thrombocytes (number µl ⁻¹)	19.000 ± 6577 ^a	12.682 ± 3856 ^b	30.8747 ± 8073 ^c	27.230 ± 14599 ^{ac}
Leukocytes (number µl ⁻¹)	48.877 ± 9706 ^a	30.683 ± 9667 ^b	43.235 ± 12115 ^a	42.062 ± 12675 ^a
Lymphocytes (number µl ⁻¹)	20.938 ± 5151 ^a	11.762 ± 5059 ^b	13.209 ± 6887 ^b	11.193 ± 4164 ^b
Monocytes (number µl ⁻¹)	18.397 ± 6083 ^{ac}	11.040 ± 3980 ^b	15.359 ± 4826 ^a	22.330 ± 8524 ^c
Neutrophils (number µl ⁻¹)	4589 ± 2927 ^a	4611 ± 3330 ^a	9646 ± 5586 ^b	6.565 ± 3654 ^{ab}
Eosinophils (number µl ⁻¹)	3269 ± 2494 ^a	2.029 ± 943 ^{ab}	3349 ± 2225 ^a	1.380 ± 721 ^b
PAS-LG (number µl ⁻¹)	1802 ± 9618 ^a	1431 ± 1042 ^{ab}	1671 ± 1163 ^a	743 ± 334 ^b

Means followed by different letters in the different rows indicate differences between treatments by the Dunn test (p <0.05). Values expressed as mean ± standard deviation. PAS-LG: PAS-positive granular leukocytes.

Table 5. Number of hepatocytes, hepatic glycogen and hepatosomatic index (HSI) of *Colossoma macropomum* fed on different concentrations of Homeopatila 100® for 60 days.

Treatments (ml/kg)	No. hepatocytes	Glycogen (%)	HSI (%)
Control	247.8 ± 38.1 ^a	19.5 ± 2.7 ^a	2.17 ± 0.15 ^{ab}
20	251.8 ± 38.1 ^a	19.8 ± 2.6 ^a	2.17 ± 0.20 ^a
40	223.3 ± 34.1 ^b	19.4 ± 1.6 ^a	1.99 ± 0.34 ^b
60	246.9 ± 25.7 ^a	19.0 ± 2.2 ^a	2.05 ± 0.29 ^{ab}

Means followed by different letters in the same column indicate differences by the Dunn test (p <0.05). Values expressed as mean ± standard deviation.

fish treated with 40 ml of Homeopatila 100®/kg of feed when compared to the other treatments, but the hepatosomatic index was lower than that in fish treated with 20 ml of homeopathic product (Table 5).

There was no morphological change in the liver of fish fed on diets containing homeopathic product and controls. Fish treated with 20 ml Homeopatila 100®/kg diet showed diffuse melanomacrophage centers in the hepatocytes and blood vessels, while in fish kept in 40 ml homeopathic product, such structures were only observed in blood vessels. However, in control fish and with 60 ml homeopathic product melanomacrophage centers were observed (Figure 2).

DISCUSSION

In Tambaqui fed control diet 20 ml of hydroalcoholic solution, 20, 40 and 60 ml of Homeopatila100® there was no mortality for 60 days. Such concentrations of

Homeopatila 100® when added to the diet of Nile tilapia increased the survival of fingerlings for 60 days (Siena et al., 2010; Braccini et al., 2013). 20 and 40 ml Homeopatila 100®/kg did not affect the performance parameters of tambaqui similarly to that described for Nile tilapia fed the same concentrations of that homeopathic product in commercial feed (Siena et al., 2010). This homeopathic product had no benefic effect on feed conversion of tambaqui, since the best results were observed in the control group, unlike the expected, as it improved feed conversion in Nile tilapia fed on diets containing 40 ml of Homeopatila 100®/kg feed (Siena et al., 2010; Braccini et al., 2013). In addition, feeding on 60 ml Homeopatila 100® reduced weight gain, mean daily gain, specific growth rate and increased feed conversion of tambaqui. Moreover, this concentration did not affect apparent feed conversion, weight and length of Nile tilapia (Siena et al., 2010; Braccini et al., 2013) kept under similar conditions to the present study.

The condition factor, a quantitative indicator of fish's

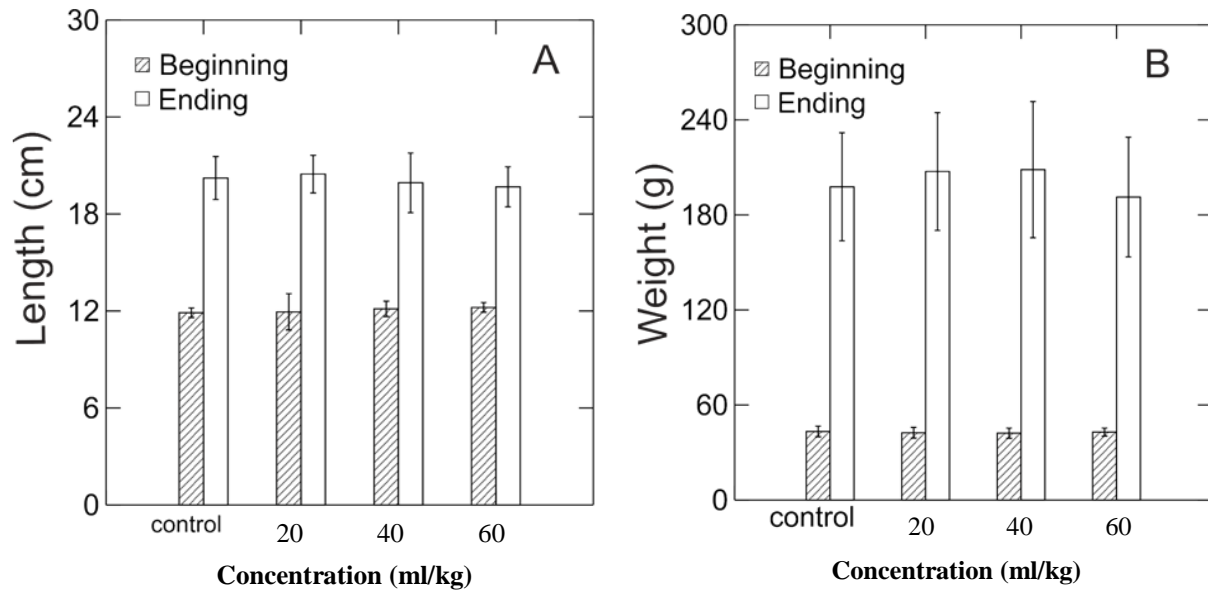


Figure 1. Initial and final length (A) and initial and final body weight (B) of *Colossoma macropomum* fed during 60 days with different concentrations of Homeopatila 100[®]. Values expressed as mean \pm standard deviation.

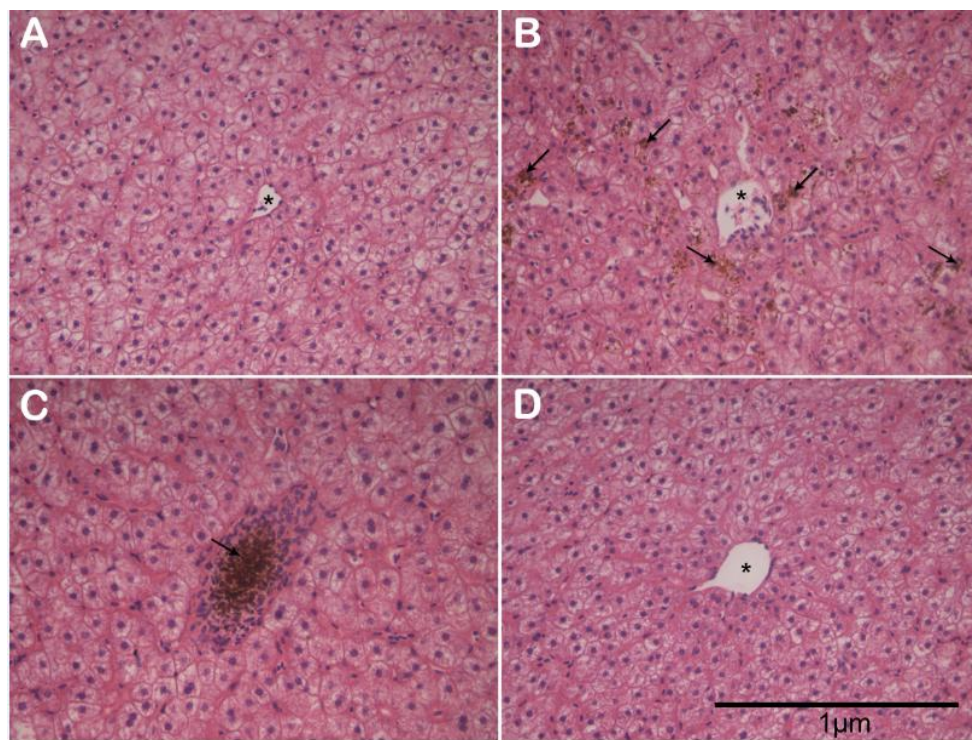


Figure 2. Liver morphology of *Colossoma macropomum* fed 0 mL of Homeopatila 100[®]/kg. (A) 20 mL, (B) 40 mL, (C) and 60 mL (D), highlighting the presence of melanomacrophage centers (arrows). *Indicate blood vessel.

body condition used to evaluate the different feeding conditions, interference of population density and other environmental conditions (Le-Cren, 1951; Guidelli et al.,

2011) showed no difference among treatments of Homeopatila 100[®] in tambaqui which was similar to that found in Nile tilapia fed this same homeopathic product

and at the same concentrations (Valentim-Zabott et al., 2008).

No difference in mean daily gain and specific growth rate was observed in tambaqui (42.7 ± 3.1 g) fed diet containing 32% crude protein and 20 ml of hydroalcoholic solution (control), 20 and 40 ml of Homeopatila 100[®]. Such growth performance parameters were higher than those described by Lemos et al. (2012) for the same fish (7.7 ± 0.2 g) when fed diets containing 26% crude protein; but the feed conversion was better. Moreover, the weight gain was similar to that reported by Pereira-Junior et al. (2013), for tambaqui (6.6 ± 0.1 g) fed on diets containing 38.3% crude protein, but apparent feed conversion was also lower. However, such differences are attributed to the initial fish size, which were larger in this study, and also to the different levels of protein used in the diet. Therefore, these results indicate that, during growth, tambaqui seems to have better feed conversion than at the beginning of fattening.

The gills of tambaqui fed diets containing 20 ml of hydroalcoholic solution (controls), 20, 40 and 60 ml Homeopatila 100[®]/kg diet were found infected by *I. multifiliis*, *P. pillulare* and four monogenean species (*A. spathulatus*, *N. janauachensis*, *M. boegeri* and *L. brinkimanni*), but *I. multifiliis* was the most abundant parasite and *P. pillulare* was only found in fish treated with 60 ml of this product. However, with the use of homeopathic product only *L. brinkimanni* showed reduction in prevalence, because regardless of treatment, all fish were parasitized by *I. multifiliis* and monogenean species, but the abundance of these parasites was higher in fish treated with 60 ml of product homeopathic. In addition, there was no difference in the prevalence of *Trichodina* sp. and Gyrodactylidae species as well as the mean intensity of parasites for Nile tilapia maintained for 60 days with diets containing these same concentrations of Homeopatila 100[®] (Braccini et al., 2013).

Tambaqui fed 60 ml Homeopatila 100[®] had lower growth performance and hence were the most parasitized by monogeneans and *I. multifiliis*, since the latter is an opportunistic protozoan. Moreover, infection with *P. pillulare*, another opportunistic protozoan, occurred only in those fish. However, for goats using homeopathic products, the number of eggs of gastrointestinal helminthes was reduced (Neves et al., 2012), but for sheep these drugs had no antiparasitic efficacy (Cavalcante et al., 2007; Signoretti et al., 2008), as has occurred in fish in the present study. Although the use of homeopathic products do not always have efficacy against helminthes, homeopathy can help to reduce the effects of parasitic infections, balancing the host-parasite relationship (Cavalcante et al., 2007; Signoretti et al., 2008).

In this study, only fish fed diets containing 20 and 60 ml Homeopatila 100[®] had higher hematocrit and higher MCV, while the levels of plasma protein, hemoglobin concentration, hematocrit, red blood cell count and MCHC

were not affected by treatment with the homeopathic product. However, Nile tilapia also fed with Homeopatila 100[®] showed reduction in plasma levels of cortisol, glucose, hematocrit, hemoglobin, red blood cell count and MCHC (Vargas and Ribeiro, 2009), then featuring a macrocytic-hypochromic anemia. On the other hand, no sign of anemia occurred in all fish of this study, but there was a higher concentration of glucose in fish of the highest concentrations of Homeopatila 100[®], showing signs of stress. However, fish treated with 60 ml of Homeopatila 100[®] had higher parasitism, caused possibly by stress (Tavares-Dias et al., 2009b).

The thrombocytes are multifunctional cells of fish, since they primarily participate in the coagulation process and secondly, assist the defense mechanism (Ranzani-Paiva et al., 2013; Santos and Tavares-Dias, 2011), thus being in constant movement between the hematopoietic organs and circulation. Reduction in the number of these blood cells may indicate a hemostatic disorder. In this study, fish fed 20 ml Homeopatila 100[®] showed reduced number of thrombocytes, while those fed 40 ml showed an increase. Vargas and Ribeiro (2009) reported increased percentage of thrombocytes for Nile tilapia fed with this same homeopathic product.

In fish, the leukocyte count is an important tool to infer the state of health and immune system because of the many functions of these cells. Lymphocytes are white blood cells involved in a variety of immune functions such as immunoglobulin production and modulation of defense. Neutrophils are the first phagocytic leukocytes in response to infection. Monocytes and PAS-LG are phagocytes that perform migration to the inflammatory site during infectious processes. Eosinophils are white blood cells that may participate in the defense process against parasites (Ranzani-Paiva et al., 2013; Santos and Tavares-Dias, 2011). In *C. macropomum*, 20 ml of Homeopatila 100[®] caused leukopenia due to lymphocytopenia and monocytopenia, while 40 ml of this product led to a Neutrophilia accompanied by lymphocytopenia. However, treatment with 60 ml of Homeopatila 100[®] resulted in monocytophilia accompanied by lymphocytopenia, eosinopenia and reduced number of LG-PAS. Similarly, the use of Homeopatila 100[®] also caused a reduction in the percentage of lymphocytes and eosinophils in Nile tilapia (Vargas and Ribeiro, 2009). Neutrophilia has been reported to fish with parasitic infections and sometimes can be accompanied by lymphocytopenia, depending on the stage of infection and effects of stress caused by parasitism (Santos et al., 2011).

In fish, the liver is a hematopoietic organ, but also stores large amount of glycogen and fat, then such reserves can influence its weight, also interfering with the hepatosomatic index (Tavares-Dias et al., 2000; Barbosa et al., 2011). In *C. macropomum*, 60 days of feeding on diets containing different concentrations of Homeopatila 100[®] did not affect the Hepatosomatic index and per-

centage of hepatic glycogen, but 40 mL reduced the amount of hepatocytes. However, in *O. niloticus* treated with Homeopatila 100[®], there was decreased amount of lipid inclusion in the liver and consequently in the HSI, but it has been reported to increase on hepatocytes number (Siena et al., 2010; Vargas and Ribeiro, 2009).

The liver also controls many vital functions as it has important role in physiology and immunity. It has perisinusoidal cells of the reticulum-endothelial system that, when present, have a phagocytic function, because they are a type of macrophage known as melanomacrophages (Bombonato et al., 2007). The melanomacrophages have several functions in fish, such as phagocytosis or pathogens, antigen processing in the immune response, destruction, detoxication or recycling of endogenous and exogenous materials, deposits of metabolites of dead cells, including red blood cells, as well as response to different antigens (Campos et al., 2008; Manrique et al., 2014). *Colossoma macropomum* maintained with 20 ml Homeopatila 100[®]/kg diet showed diffuse melanomacrophages centers in hepatocytes, but in fish kept with 40 ml of this homeopathic product, such structures were restricted to vascular channels (Campos et al., 2008).

Conclusion

For *C. macropomum* the Homeopatila 100[®] did not reduce the infections of parasites in the gills, but showed a relative improvement in blood parameters of fish fed on 40 ml. As the Homeopatila 100[®] has not improved the performance of the fish; therefore, the use of this homeopathic product is an additional cost in fish production for the fish farm. However, this homeopathic product was prepared in principle aiming at requirements of Nile tilapia, African fish with different biology from *C. macropomum*, an Amazonian fish.

ACKNOWLEDGEMENTS

This research followed the principles adopted by the Colégio Brasileiro de Experimentação Animal (COBEA). The authors are indebted to CNPq for the financial support (# 472054/2013-9) and for the PQ scholarship awarded to Dr. Tavares-Dias, M.

Conflict of interest

Authors declare that there are no conflicts of interest

REFERENCES

Araujo-Lima CARM, Gomes LC (2005). Tambaqui (*Colossoma macropomum*). In: Baldissarroto B, Gomes LC (eds). Espécies

- nativas para piscicultura no Brasil. 2ª ed. Santa Maria: UFSM. pp. 67-104.
- Barbosa MC, Jatobá A, Vieira FN (2011). Cultivation of juvenile fat snook (*Centropomus parallelus* Poey, 1860) fed probiotic in laboratory conditions. Braz. Arch. Biol. Technol. 54:795-801.
- Behmer OA, Tolosa EMC, Freitas NETO AG (1976). Manual de técnicas para histologia normal e patológica. São Paulo: Edusp/Edart p 256.
- Beçak W, Paulete J (1976). Técnicas de citologia e histologia. Rio de Janeiro: Livros técnicos Científicos. p 574.
- Bombonato MTS, Rochel SS, Vicentini CA, Vicentini IBF (2007). Estudo morfológico do tecido hepático de *Leporinus macrocephalus*. Acta Sci. Biol. Sci. 29:81-85.
- Braccini GL, Natali MRM, Ribeiro RP, Mori RH, Riggo R, Oliveira CAL, Hildebrandt JF, Vargas L (2013). Morpho-functional response of Nile tilapia (*Oreochromis niloticus*) to a homeopathic complex. Homeopathy 102:233-241.
- Bush AO, Lafferty KD, Lotz JM, Shostak W (1997). Parasitology meets ecology on its own terms: Margolis et al. revisited. J. Parasitol. 83:575-583.
- Campos CM, Moraes JRE, Moraes FR (2008). Histopatologia de fígado, rim e baço de *Piaractus mesopotamicus*, *Prochilodus lineatus* e *Pseudoplatystoma fasciatum* parasitados por myxosporídios, capturados no Rio Aquidauana, Mato Grosso do Sul, Brasil. Rev. Bras. Parasitol. Vet. 17:200-2005.
- Cavalcante ASR, Almeida MAO, Dias AVS (2007). Efeito de medicamentos homeopáticos no número de ovos de nematoides nas fezes (OPG) e no ganho de peso em ovinos. Rev. Bras. Prod. Anim. 8:162-169.
- Eiras JC, Takemoto RM, Pavanelli GC (2006). Métodos de estudo e técnicas laboratoriais em parasitologia de peixes. 2ª d. Maringá: Eduem. p 199.
- Fracalossi DM, Rodrigues APO, Silva TSC, Cyrino JEP (2013). Técnicas experimentais em nutrição de peixes. In: Fracalossi DM, CyrinoJEP (eds). Nutriaqua. 1ª ed. Florianópolis: Aquabio. pp. 37-63.
- Guidelli G, Tavechio WLG, Takemoto RM, Pavanelli GC (2011). Relative condition factor and parasitism in anostomid fishes from the floodplain of the upper Paraná River, Brazil. Vet. Parasitol. 177: 145 - 151.
- Inoue LAKA, Bojijn kCL, Ribeiro PT, Silva AMD, Affonso EG (2011). Avaliação de respostas metabólicas do tambaqui exposto ao eugenol em banhos anestésicos. Acta Amaz. 41:327-332.
- Le-Cren ED (1951). The length-weight relationship and seasonal cycle in gonadal weight and condition in the perch (*Percafluviatilis*). J. Anim. Ecol. 20:201-219.
- Lemos MVA, Guimaraes IG, Miranda EC (2011). Farelo de coco em dietas para tambaqui. Rev. Bras. Prod. Anim. 12:188-198.
- Manrique WG, Claudiano GS, Petrillo TR, Pardi MC, Pereira FMA, Belo MAA, Moraes JRE, Moraes FR (2014). Response of splenic melanomacrophage centers of *Oreochromis niloticus* (Linnaeus, 1758) to inflammatory stimuli by BCG and foreign bodies. J. Appl. Ichthyol. 30:1-6.
- Ministério da Pesca e Aquicultura-MPA (2013). Boletim estatístico da pesca e aquicultura 2011. Brasília, DF. p 60.
- Neves HH, Hotzel MJ, Honorato LA, Fonseca CEM, Mata MGF, Silva JB (2012). Controle de verminose gastrointestinal em caprinos utilizando preparados homeopáticos. Rev. Bras. Agroecol. 7:45-151.
- Oliveira ACB, Miranda EC, Correa R (2013). Exigências nutricionais e alimentação de tambaqui. In: Fracalossi DM, CyrinoJEP (eds). Nutriaqua. 1ª ed. Florianópolis: Aquabio. pp 231-240.
- Pereira-Junior GP, Pereira EMO, Filho MP, Barbosa OS, Shimoda E, Brandão LV (2013). Desempenho produtivo de juvenis de tambaqui (*Colossoma macropomum* Cuvier, 1818) alimentados com ração contendo farinha de cruzeira de mandioca (*Manihotesculenta*, Crantz) em substituição ao milho (*Zeamays*). Acta Amaz. 43: 217-226.
- Ranzani-Paiva MJT, Pádua SB, Tavares-Dias M, Egami MI (2013). Métodos para análises hematológicas em peixes. Maringá: Eduem. p 140.
- Rocha CMC, Resende EK, Routledge EAB, Lundstedt LM (2013). Avanços na pesquisa e no desenvolvimento da aquicultura brasileira. Pesq. Agropec. Bras. 48(8):4-6.
- Santos RBS, Tavares-Dias M (2011). Células sanguíneas e resposta

- hematológica de *Oxydorasniger* (Pisces, Doradidae) oriundos da bacia do médio Rio Solimões, estado do Amazonas (Brasil), naturalmente parasitados. Bol. Inst. Pesca. 36:283-292.
- Santos EF, Tavares-Dias M, Pinheiro DP, Neves LR, Marinho RGB, Dias MKR (2013). Fauna parasitária de tambaqui *Colossoma macropomum* (Characidae) cultivado em tanque-rede no estado do Amapá, Amazônia oriental. Acta Amaz. 43:107-114.
- Siena CE, Natali MRM, Braccini GL, Oliveira AC, Ribeiro RP, Vargas L (2010). Efeito de um núcleo homeopático homeopatia 100[®] na eficiência produtiva em alevinos revertidos de tilápia do Nilo (*Oreochromis niloticus*). Semina: Ciên. Agric. 31:985-994.
- Signoretti RD, Veríssimo CJ, Souza FHM, Garcia TS, Oliveira EM, Souza KG, Mourão GB (2008). Desempenho e infestação por parasitos em machos leiteiros suplementados com sal proteinado com ou sem os medicamentos homeopáticos. Rev. Bras. Parasitol. Vet. 17:40-44.
- Tavares-Dias M, Sandrin EFS, Campos-Filho E (1999). Características hematológicas do tambaqui *Colossoma macropomum* Cuvier (Osteichthyes: Characidae) em sistema de monocultivo intensivo. II. Leucócitos. Rev. Bras. Zool. 16:175-84.
- Tavares-Dias M, Martins ML, Moraes FR (2000). Relação hepatossomática em peixes teleósteos de cultivo intensivo. Rev. Bras. Zool. 17:273-281.
- Tavares-Dias M, Ishikawa MM, Martins ML, Sateke F, Hisano H, Pádua SB, Jerônimo Sá ARS (2009b). Hematologia: ferramenta para o monitoramento do estado de saúde dos peixes em cultivo. In: Netto AS, Mariano WS, Soria SFP. Tópicos Especiais em Saúde e Criação Animal. São Carlos: Pedro & João. pp 43-80.
- Tavares-Dias M, Araújo CSO, Porto SMA, Viana GM, Monteiro PC (2013). Sanidade de tambaqui *Colossoma macropomum* nas fases de larvicultura e alevinagem. Macapá: Embrapa Amapá. p 42.
- Valentim-Zabott M, Vargas L, Ribeiro RP, Piau JRR, Torres MBA, Ronnau M, Souza JC (2008). Effects of a homeopathic complex in Nile tilapia (*Oreochromis niloticus* Linnaeus) on performance, sexual proportion and histology. Homeopathy 97:190-195.
- Vargas L, Ribeiro RP (2009). Homeopatia populacional em tilápias do Nilo *Oreochromis niloticus*. In: Tavares-Dias M. (ed.). Manejo e sanidade de peixes em cultivo. Macapá: Embrapa Amapá. pp.106-131.

Full Length Research Paper

***In vitro* evaluation of the antibacterial activities of the methanol, aqueous and n-hexane extracts of *Ocimum lamiifolium* from Ethiopia**

Destaw Damtie and Yalemtehay Mekonnen*

Cellular and Molecular Biology Department, Addis Ababa University, Ethiopia.

Received 6 August, 2014; Accepted 26 January, 2015

Ocimum lamiifolium (local name Dama Kesse, Amharic) is a medicinal plant in Ethiopia. Its leaves are squeezed and sniffed to treat coughs and colds. They are also used to treat eye infections and to stop nose bleedings. In the present study, leaves of *O. lamiifolium* were collected from their growing habitats. Dried leaf powders were extracted using methanol, distilled water and n-hexane. 25, 50, and 100 mg/ml doses of the extracts made in Tween 80 (2%) were screened for their antimicrobial activities against *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa* and *Shigella boydii* using disk diffusion assay. The inhibition zones due to the methanolic extract ranged from 0 (in *S. aureus* due to 25 mg/ml) to 12 mm (in *E. coli* due to 100 mg/ml). Inhibition zones due to the aqueous extract ranged from 8 mm in *S. aureus* and *S. boydii* to 12 mm in *S. boydii* at concentrations of 25 and 100 mg/ml, respectively. The n-hexane extract at 25 mg/ml resulted in inhibition zone that ranges from 7 mm (against *S. aureus*) to 11 mm (against *E. coli*) at 50 and 100 mg/ml doses. The minimum inhibitory concentration of *S. boydii* and *E. coli* was 10 mg/ml due to all the extracts. The minimum inhibitory concentrations on *S. aureus* were 10, 20 and 50 mg/ml due to the aqueous, n-hexane and methanolic extracts, respectively. *P. aeruginosa* was minimally inhibited at 10 mg/ml due to the methanol and aqueous extracts and 15 mg/ml due to the n-hexane extract. The methanol, aqueous, and n-hexane extracts of *O. lamiifolium* leaf extracts inhibited the test bacteria with significantly higher levels of inhibition zones than the negative control (T80). The positive controls (Tetracycline and Chloramphenicol) also showed significantly higher inhibition zones than the 100 mg/ml concentration of the extracts and T80 except that Chloramphenicol failed to inhibit *S. aureus* and *P. aeruginosa*. However, combination of Chloramphenicol with plant extracts raised their inhibition zones from zero to 23 and 25 mm in *S. aureus* and *P. aeruginosa*, respectively.

Key words: *Ocimum lamiifolium*, antibacterial activity, methanolic extract, aqueous extract, n-hexane extract.

INTRODUCTION

The genus *Ocimum* (Lamiaceae) consists of about 30

species distributed in the tropics and subtropics of the

*Corresponding author. E-mail: zegades529@yahoo.com; yalemtmek55@gmail.com.

Author(s) agree that this article remain permanently open access under the terms of the [Creative Commons Attribution License 4.0 International License](http://creativecommons.org/licenses/by/4.0/)

Old and New worlds, with some species cultivated in temperate areas. *Ocimum lamiifolium* Hochst.ex Benth(local name Tossign, Amharic) is mostly found in clearings and edges of primary and secondary mountain forests and bushlands, tall grasslands, abandoned fields, at altitudes between 1200 and 2900 m. Traditionally, the fresh leaves are squeezed and the juice is sniffed to treat cough and cold. The juice is also used as eye rinse to treat eye infections. At the same time, the crushed leaves are put in the nostrils to stop nose bleeding (Asfaw and Demissew, 2009).

Biologically, different extracts of the genus *Ocimum* are known for their antibacterial (Nakamura et al., 1999; Nascimento et al., 2000; Adebolu and Oladimeji, 2005; Adiguzel et al., 2005; Ahmad and Aqil, 2007; Goyal and Kaushik, 2011; Patil et al., 2011; Sneha et al., 2011; Prasannabalaji et al., 2012), antifungal (Amadioha, 2001), and antioxidant (Hakkim et al., 2008) activities. *O. lamiifolium* extracts are also known to have antibacterial, antifungal, insecticidal and insect repellent (Dagne, 2009), antiinflammatory (Kashyap et al., 2011) activities. The chemical composition of essential oils of six *Ocimum* species from East Africa including *O. lamiifolium* were majorly phenyl propane derivatives or terpenoids, including methyl eugenol, 1, 8-cineole, camphor, bornyl acetate, germacrene-D, E-myroxide, germacrene-B, caryophyllene oxide and p-cymene (Kashyap et al., 2011). In another study by Tchoumboungang et al. (2014), 85.7% of *O. lamiifolium* essential oils were monoterpenes [sabinene (33.8%), (Z)- β -ocimene (17.2%), terpinen-4-ol (8.4%) and others] and sesquiterpenes (8.7%) [β -caryophyllene (5.6%), germacrene D (1.1%), (E)- β -farnesene (1%) and others]. Sabinene is known to have inhibition effects against Gram negative and Gram positive bacteria (Wiar, 2006; Uinoiseau et al., 2010). In addition, eugenol, a component of *Ocimum* has antibacterial and antihelmintic activities (Adebolu and Oladimeji, 2005). Components of *Ocimum basilicum* like apigenin, linalool and ursolic acid, exhibit a broad spectrum of antiviral activity and are used as remedies for treating disorders such as viral ocular, respiratory and hepatic infections (Chiang et al., 2005). The aim of the present study, however, was to test the antibacterial activities of the methanol, aqueous, and n-hexane extracts of the leaves of *O. lamiifolium*.

MATERIALS AND METHODS

Dried and powdered leaves of *O. lamiifolium*, methanol (Reagent chemical Services Ltd., United Kingdom), n-hexane (Uni-Chem Chemical Reagents), nutrient agar (Oxoid LTD., Basingstoke, Hampshire, England), Muller-Hinton agar (Oxoid LTD., Basingstoke, Hampshire, England), sulfuric acid (SDFCL Fine Chemical Ltd., Mumbai, India), Tween 80 (Uni-Chem Chemical Reagents), sodium chloride (Nike Chemical, India), cotton swab (Nataso, India), tetracycline (Oxoid Ltd., United Kingdom), chloramphenicol (Oxoid Ltd., United Kingdom), barium chloride (BDH Chemicals Ltd. Poole, England), an autoclave (Express autoclave, Dixons surgical Ltd.), petri dishes, and distilled water (Biomedical Laboratory, Addis

Ababa University, Ethiopia).

Plant collection and identification

O. lamiifolium leaves were collected from their natural habitats Central and North East Ethiopia. The plants were not flowering during the period of collection. The collected specimens were authenticated by botanists from the National Herbarium of Addis Ababa University and voucher specimens were deposited at the same herbarium of Addis Ababa University.

Extraction of plant

Collected leaves of *O. lamiifolium* were washed by distilled water and subjected to shade drying at 25°C. Then the dried leaves were pulverized to get coarse powder. 100 g of the powder was added to 1 L (1:10, w:v) of three solvent types, namely, methanol (absolute), n-hexane (absolute), and distilled water and each mixture was shaken for 48 h at 120 rotations/min. The solutions were filtered by Whatman No. 1 filter paper. Finally, the methanol and hexane extracts were concentrated under vacuum in a rotary evaporator (Büchi Laboratoriums-Tchnik AG CH-9230 Flawil/Schweiz) to give gummy residues and the aqueous extracts using a lyophilizer (Bioblock Scientific, Illkirch Cedex, France). The crude extracts were then weighed and the yield of the each extract was calculated as 17.8, 12 and 6.7% (methanol, aqueous and n-hexane extract, respectively).

Bacterial strains

Clinical isolates of *Staphylococcus aureus*, *Shigella boydii*, *Escherichia coli*, and *Pseudomonas aeruginosa* were obtained from the Ethiopian Public Health Institute (EPHI). These isolates were screened for their susceptibility towards different doses of the different extracts of *O. lamiifolium* as well as two standard antibiotics [Tetracycline (30 μ g/disk) and Chloramphenicol (30 μ g/disk)]. In order to perform the antimicrobial screening, the bacterial isolates were cultured overnight at 37°C on Nutrient Agar medium. Colonies collected from each 24 h old bacterial culture were diluted in sterile saline and the optical density was adjusted in comparison with 0.5 McFarland' scale to prepare a standardized inoculum (1.5×10^8 cfu/ml). The bacteria from saline solutions were spread on Müller Hinton Agar plates using sterile cotton swabs.

The paper disc diffusion technique was applied to determine the antimicrobial activities of the tested plant extracts. Sterile paper discs (5 mm in diameter) immersed in stock solutions containing 25, 50 and 100 mg/ml prepared in 2% Tween 80 of plant extracts were placed on the surface of inoculated Nutrient Agar plates. Plates were then incubated for 24 h at 37°C, and diameters of the inhibition zones were recorded. All assays were applied in triplicates and the results are given as means \pm standard error of the mean.

Determination of minimum inhibitory concentration (MIC)

MIC is the lowest concentration of an antimicrobial that inhibits the visible growth of microorganisms after overnight incubation (Yilmaz, 2012). MICs were defined as the lowest concentration of the aqueous, methanol and n-hexane extracts of *O. lamiifolium* inhibiting visible growth of the bacteria. On the other hand, the MBC was defined as the lowest concentration of the extracts of *O. lamiifolium* required to kill all the test bacteria (Yilmaz, 2012). The MIC was determined using agar dilution method which is described by the European Committee for Antimicrobial Susceptibility Testing (EUCAST) of the European Society of Clinical Microbiology and

Table 1. Minimum inhibitory concentrations (MIC) of *Ocimum lamiifolium* leaf extracts.

Bacterial species	Gram type	MIC (mg/ml)		
		Methanol extract	Aqueous extract	n-Hexane extract
<i>S. boydii</i>	-	10	10	10
<i>S. aureus</i>	+	50	10	20
<i>E. coli</i>	-	10	10	10
<i>P. aeruginosa</i>	-	10	10	15

Infectious Diseases (ESCMID, 2000). The following procedure was followed to determine the MIC; 20 ml agar was used in 9-cm Petri dishes for agar dilution. Nineteen-milliliters molten agar was added to 1 ml of each plant extract to make the total volume 20 ml. Müller Hinton agar was prepared as recommended by the manufacturer. The sterilized agar was set to cool to 50°C in a water-bath. Extracts of *O. lamiifolium* were prepared into doses of 5, 10, 15, 20, 25, 50 and 100 mg/ml in 25 to 30 ml containers. Nineteen-milliliters of molten agar was added to each container and mixed thoroughly, and finally poured into pre-labeled sterile Petri dishes on a level surface. The plates were allowed to dry at room temperature so that no drops of moisture remain on the surface of the agar.

Bacterial suspensions were prepared in 0.85% normal saline and were standardized by 0.5 McFarland standards to 1.5×10^8 colony forming units (CFU)/ml. The inocula were inoculated on the dry plates. The inoculum spots were then allowed to dry at room temperature before inverting the plates for incubation. Finally, the plates were incubated at 37°C in air for 18 h. The MIC (the lowest concentration of the extracts that completely inhibited visible growth) was judged by the naked eye.

Determination of susceptibility test of bacteria towards standard antibiotics and their combinations with *O. lamiifolium* extracts

Susceptibility of the test bacteria towards Chloramphenicol, Tetracycline and their combinations with the leaf extracts of *O. lamiifolium* was determined according to the classification indicated by Bauer et al. (1966). Based on this literature, inhibition zones due to Chloramphenicol (30 µg) can be classified as resistant (≤ 12 mm), intermediate (13 to 17 mm), and sensitive (≥ 18 mm) and zones of inhibition for Tetracycline (30 µg) are interpreted as resistant (≤ 14 mm), intermediate (15 to 18 mm), and sensitive (≥ 19 mm).

RESULTS

Antibacterial activities of *O. lamiifolium* leaf extracts

The methanol extract inhibited the test bacteria in a dose dependent manner (Figure 1). At 25 mg/ml, it did not inhibit *S. aureus* while the rest bacteria were inhibited by this dose with inhibition zones just below 10 mm. The 50 mg/ml concentration of the methanol extract on the other hand inhibited all the bacteria with mean inhibition zones ranging from 6 mm (*S. aureus*) to over 10 mm (*E. coli*). At 100 mg/ml concentration, the methanol extract inhibited three of the test bacteria with mean inhibition zones above 10 mm and *S. aureus* with mean inhibition zone close to 10 mm. The antibacterial activity of the methanol

extract was generally lower than that of Tetracycline and Chloramphenicol. However, it was better than Chloramphenicol in inhibiting *S. aureus* and *S. boydii*. The aqueous extract inhibited all the test bacteria at 25, 50 and 100 mg/ml doses minimally inhibiting *S. aureus* and *E. coli* each with mean inhibition zones of 8 mm at 25 mg/ml dose and maximally *S. boydii* (12 mm) at 100 mg/ml. The aqueous extracts too were generally less effective than Tetracycline and Chloramphenicol although Chloramphenicol resistant strains (*S. aureus* and *P. aeruginosa*) were sensitive to these extracts. Like that of the aqueous extract, the n-hexane extract inhibited all the test bacteria at the three dose levels, *S. aureus* being inhibited minimally (7 mm) at 25 mg/ml and *E. coli* being inhibited maximally (11 mm) at 100 mg/ml. In general, the trend of inhibition of the test bacteria by the three extracts of *O. lamiifolium* showed that the aqueous extract is the best followed by its methanol and n-hexane extracts, respectively.

Determination of the MIC

The MIC concentrations of *O. lamiifolium* leaf extracts ranged from 10 to 50 mg/ml (Table 1). The 50 mg/ml concentration of its methanol extract inhibited all the bacteria and its 10 mg/ml inhibited 75% of them. The aqueous extract, on the other hand, inhibited all the bacteria at a concentration of 10 mg/ml and the n-hexane extract inhibited the microorganisms with a range of MICs from 10 to 20 mg/ml of which the 20 mg/ml inhibited the entire, 15 mg/ml inhibited 75%, and 10 mg/ml inhibited 50 percent of them.

Antibacterial effects of the combinations of Tetracycline (30 µg/ml) and Chloramphenicol (30 µg/ml) with *O. lamiifolium* leaf extracts at 100 mg/ml

Chloramphenicol (30 µg) resulted in inhibition zones of 31 and 33 mms in *S. boydii* and *E. coli*, respectively (Figure 2). On the contrary, it did not inhibit *S. aureus* and *P. aeruginosa*. Tetracycline (30 µg) inhibited *S. aureus* with inhibition zone of 11 mm and the rest bacteria with inhibition zones above 19 mm. Combination of these antibiotics to *O. lamiifolium* extracts at a dose of 100

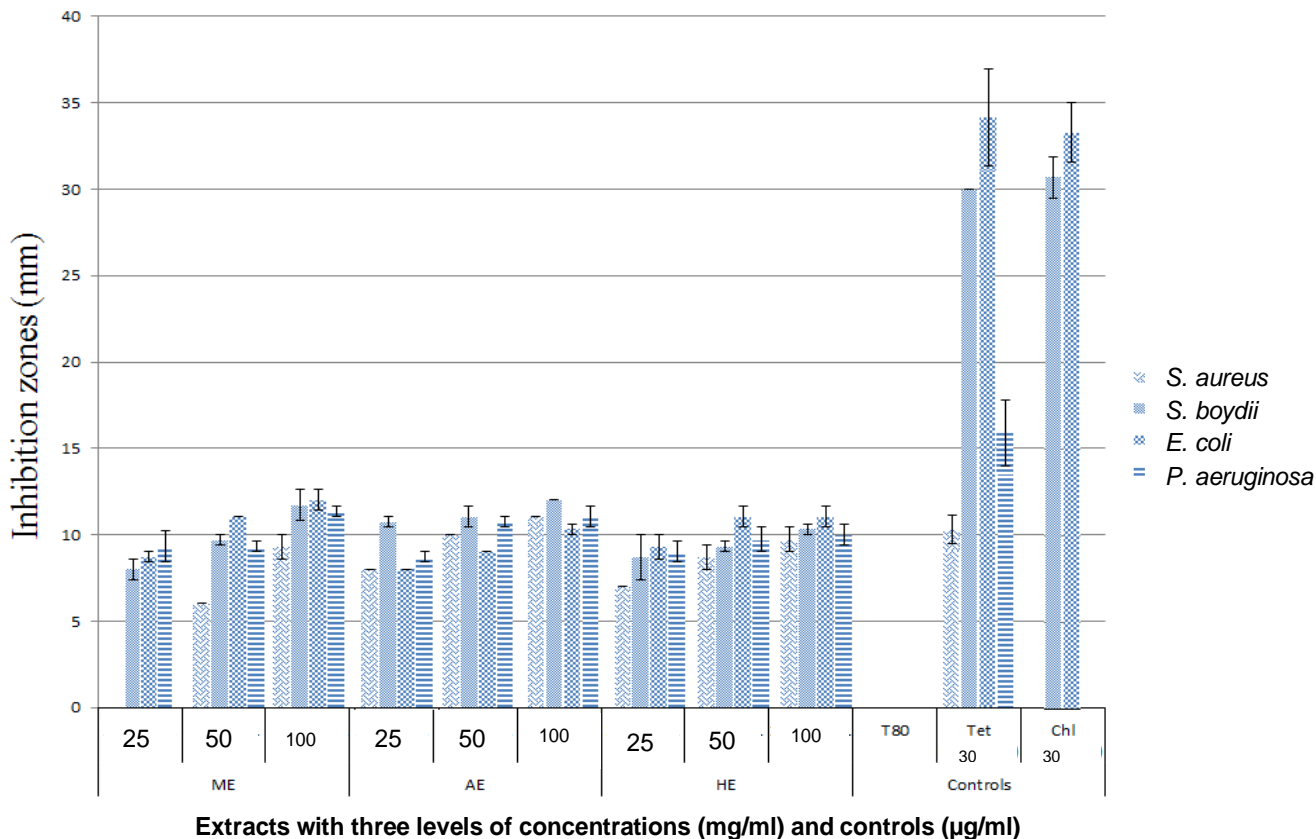


Figure 1. Inhibition zone (mm) of *O. lamiifolium* leaf extracts against Gram positive and negative bacteria. Tet: Tetracycline; Chl: Chloramphenicol; T80: Tween 80; ME: Methanol extract; AE: Aqueous extract; HE: n-hexane extract.

mg/ml, however, increased the inhibition zones of the test bacteria. Inhibition of *S. boydii* and *E. coli* due to Chloramphenicol surpassed inhibition due to the combination of Chloramphenicol and plant extracts (100 mg/ml) (Figure 2). On the contrary, inhibition of *S. aureus* and *P. aeruginosa* was found to be higher than either caused by Chloramphenicol or extracts. In the same manner, inhibition of *S. boydii* and *E. coli* by Tetracycline were higher than inhibition by combination of plant extracts (100 mg/ml) and Tetracycline while *S. aureus* and *P. aeruginosa* were more sensitive to the combinations than individual parts. Generally, combining standard drugs with plant extracts boosted the inhibition of *S. aureus* and *P. aeruginosa* than that of either the drugs or the extracts.

Differences in zones of inhibition among the methanol, aqueous, and n-hexane extracts of *Ocimum lamiifolium* at concentrations of 100 mg/ml

Differences in inhibition zones due to the different extracts of *O. lamiifolium* against the test bacteria are presented in Table 2.

DISCUSSION

Inhibition was concentration dependent in all of the bacteria with *S. boydii* being the most sensitive bacterium followed by *E. coli*, *P. aeruginosa* and *S. aureus* in decreasing order of sensitivity. Similar findings were demonstrated in a study by Gebrehiwot and Unakal (2013) where *E. coli* was the most sensitive followed by *S. aureus* and *P. aeruginosa*, respectively to the aqueous and ethanol extracts of *O. lamiifolium*. This antibacterial activity may be due to the occurrence of antibacterial active components like eugenol and sabinene in the extracts (Adebolu and Oladimeji, 2005; Wiart, 2006; Uinoiseau et al., 2010). In the present study, *S. aureus* and *P. aeruginosa* were found to be resistant to Chloramphenicol may be due to the ability of these bacteria to inactivate Chloramphenicol by enzymes coded by the *cat* genes or the ability of *P. aeruginosa* to inactivate Chloramphenicol by Chloramphenicol acetyltransferase enzyme and decreased outer membrane permeability or active efflux of this drug (Byarugaba, 2009). *S. aureus* was resistant to tetracycline may be due to its abilities like active efflux of the antibiotic and ribosome protection or modification of

Table 2. Inhibition zones of *Ocimum lamiifolium* extracts at different concentrations.

Extract	Concentration	Mean inhibition zone (mm) ± SEM			
		<i>S. aureus</i>	<i>S. boydii</i>	<i>E. coli</i>	<i>P. aeruginosa</i>
ME	25	0.00 ± 0.00	8.00 ± 0.58 ^d	8.67 ± 0.33 ^d	9.33 ± 0.88 ^{d,e}
	50	6.00 ± 0.00 ^{d,e}	9.67 ± 0.33 ^d	11.00 ± 0.00 ^d	9.33 ± 0.33 ^{d,e}
	100	9.33 ± 0.67 ^{d,e}	11.67 ± 0.88 ^d	12.00 ± 0.58 ^d	11.33 ± 0.33 ^{d,e}
AE	25	8.00 ± 0.00 ^{a,d,e}	10.67 ± 0.33 ^d	8.00 ± 0.00 ^d	8.67 ± 0.33 ^{d,e}
	50	10.00 ± 0.00 ^{a,d,e}	11.00 ± 0.58 ^d	9.00 ± 0.00 ^d	10.67 ± 0.33 ^{d,e}
	100	11.00 ± 0.00 ^{d,e}	12.00 ± 0.00 ^d	10.33 ± 0.33 ^d	11.00 ± 0.58 ^{d,e}
HE	25	7.00 ± 0.00 ^{a,d,e}	8.67 ± 1.33 ^d	9.33 ± 0.67 ^d	9.00 ± 0.58 ^{d,e}
	50	8.67 ± 0.67 ^{d,e}	9.33 ± 0.33 ^d	11.00 ± 0.58 ^d	9.67 ± 0.67 ^{d,e}
	100	9.67 ± 0.67 ^{d,e}	10.33 ± 0.33 ^d	11.00 ± 0.58 ^d	10.00 ± 0.58 ^{d,e}
Controls	Tet	[#] 11.33 ± 0.88 ^{a,b,c,d,e}	30.00 ± 0.00 ^{a,b,c,d}	34.00 ± 3.06 ^{a,b,c,d}	16.00 ± 2.08 ^{a,b,c,d,e}
	Chl	0.00 ± 0.00	30.67 ± 1.20 ^{a,b,c,d}	33.33 ± 1.67 ^{a,b,c,d}	0.00 ± 0.00
	T80	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00

ME: Methanolic extract; AE: aqueous extract; HE: n-hexane extract; ^aME; ^bAE; ^cHE; ^dT80; ^eChl; T80: Tween 80; Tet: Tetracycline (30 µg/disk); Chl: Chloramphenicol (30 µg/disk); *Significantly higher inhibition than; [#]Not significantly different from b and c at 50 mg/ml and a, b and c at 100 mg/ml extracts.

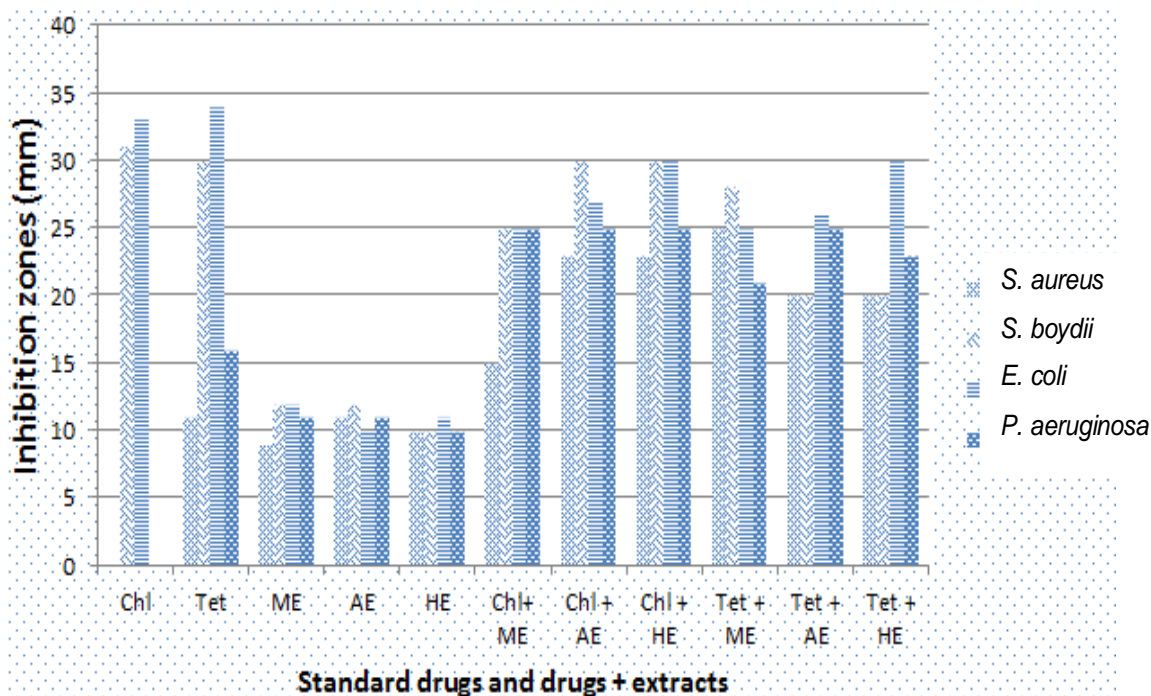


Figure 2. Antibacterial activities of the combinations of Tetracycline (30 µg) and Chloramphenicol (30 µg) with *O. lamiifolium* extracts with dose levels of 100 mg/ml. Tet: Tetracycline; Chl: Chloramphenicol; ME: Methanol extract; AE: Aqueous extract; HE: n-hexane extract.

the antibiotic (Byarugaba, 2009). All the extracts of *O. lamiifolium* showed better inhibition than the negative control (Tween 80) and were less effective than

Tetracycline and Chloramphenicol. However, the leaf extracts were more effective than Chloramphenicol against *S. aureus* and *P. aeruginosa*. On the other hand,

these extracts were less effective than Chloramphenicol against *S. boydii* and *E. coli*. Application of the aqueous extract inhibited even the most resistant bacterium (*S. aureus*) at the lowest concentration (25 mg/ml).

E. coli and *S. boydii* were the most sensitive to the extracts followed by *P. aeruginosa* and the least sensitive of all was *S. aureus* showing that the Gram negative bacteria were more sensitive to the plant extracts than the Gram positive one (*S. aureus*). The results confirmed that *O. lamiifolium* extracts are important to inhibit *S. boydii* and *E. coli*, followed by *P. aeruginosa*, and least effective against *S. aureus*. The aqueous extract of *O. lamiifolium* seem to be effective than its methanol and n-hexane extracts against these bacteria. This result clearly distinguishes the importance of the aqueous extract which contains the most effective components to inhibit bacterial growth contradicting to the finding by Goyal and Kaushik (2011) where the methanolic extract of *Ocimum sanctum* L. showed comparatively higher activity than other organic and aqueous extracts. On the other hand, this result agrees with the work of Gebrehiwot and Unakal (2013) where the aqueous extract was found to be more effective than its ethanol extract against *S. aureus*, *E. coli* and *P. aeruginosa*. Generally, differences in activities of *O. lamiifolium* extracts may be due to the differences in their chemical compositions which are determined by different factors such as climate, plant nutrition, stress (Carson and Hammer, 2011), fertilizer application (Duke, 2009), plant organs used, plant developmental stage, plant origin, chemotypes, and methods used (Zuzarte et al., 2011).

Sometimes, the effectiveness of antibiotics can be increased by coupling them with plant extracts (Kekuda, 2012). In the present study, combination of Tetracycline (30 µg/disc) to the methanol extract of *O. lamiifolium* (100 mg/ml) increased the sensitivity of *S. aureus*. On the other hand, combination of Chloramphenicol (30 µg/disc) with all the extracts increased the sensitivity of *S. aureus* and *P. aeruginosa*. The implication of this finding is that the use of plant extracts in combination with less effective antibiotics can increase the susceptibility of bacteria to these antibiotics and can be solutions to bacterial resistance to antibiotics.

The majority of the dosages of all the extracts of *O. lamiifolium* inhibited the test bacteria with inhibition zones significantly higher than that of tween 80. The positive control (Tetracycline) on the other hand inhibited all the bacteria with inhibition zones significantly higher than all the extracts except the 50 mg/ml and the 100 mg/ml concentrations of the methanol, aqueous, and n-hexane extracts of *O. lamiifolium* which resulted in inhibition zones on *S. aureus* which were not significantly different from that exerted by Tetracycline. This leads to the conclusion that the aqueous, methanol and n-hexane extracts of *O. lamiifolium* have comparable activities with Tetracycline against *S. aureus*.

Conflict of interest

The authors declare that there is no conflict of interests regarding the publication of this article.

REFERENCES

- Adebolu TT, Oladimeji SA (2005). Antimicrobial activity of leaf extracts of *Ocimum gratissimum* on selected diarrhoea causing bacteria in southwestern Nigeria. *Afr. J. Biotechnol.* 4(7):682-684
- Adiguzel A, Gulluce M, Sengul M, Ogutcu H, Sahin F, Karaman I (2005). Antimicrobial Effects of *Ocimum basilicum* (Labiatae) Extract. *Turk. J. Biol.* 29:55-160
- Ahmad I, Aqil F (2007). *In vitro* efficacy of bioactive extracts of 15 medicinal plants against ESBL-producing multidrug-resistant enteric bacteria. *Microbiol. Res.* 162:264-275.
- Amadioha AC (2001). Fungicidal effect of Some Plant Extracts against *Rhizoctonia solani* in cow pea. *Arch. Phytopathol.* 33:509-517.
- Asfaw N, Demissew S (2009). *Aromatic Plants of Ethiopia*. Addis Ababa: Shama Books. p 222.
- Bauer AW, Kirby WM, Sherris JC, Turck M (1966). Antibiotic Susceptibility Testing by a Standardized Single Disk Method. *Am. J. Clin. Pathol.* 36(3):493-496
- Byarugaba DK (2009). Mechanisms of Antimicrobial Resistance. In de J. Sosa A, Byarugaba DK, Amabile-Cuevas CF, Kariuki S, Okele IN (eds.): *Antimicrobial Resistance in Developing Countries* pp. 15-26.
- Carson CF, Hammer KA (2011). *Chemistry and Bioactivity of Essential Oils in Thormar, H* (2011) *Lipids and Essential Oils as Antimicrobial Agents*, John Wiley & Sons, Ltd, United Kingdom pp 204-238.
- Chiang LC, Ng LT, Cheng PW, Chiang W, Lin CC (2005). Antiviral Activities of Extracts and Selected Pure Constituents of *Ocimum Basilicum*. *Clin. Exp. Pharmacol. Physiol.* 32:811-816.
- Dagne E (2009). *Natural Data Base for Africa (NDA) on CD-ROM*, Addis Ababa, Ethiopia. Specie ID 302
- Duke TA (2009). *Duke's Handbook of Medicinal Plants of Latin America*. CRC Press is an imprint of Taylor & Francis Group, an Informa business, United States of America
- European Society of Clinical Microbiology and Infectious Diseases (ESCMID) (2000). *Clinical Microbiology and Infection* 6(9):509-515
- Gebrehiwot A, Unakal CG (2013). Effect of Aqueous and Ethanolic Extracts of *Ocimum lamiifolium* and *Amaranthus dubius* against Bacteria Isolated from Clinical Specimen. *Int. J. Pharm. Ind. Res.* 03(01):10-14
- Goyal R, Kaushik P (2011). *In vitro* Evaluation of Antibacterial Activity of Various Crude Leaf Extracts of Indian Sacred Plant, *Ocimum sanctum* L. *Br. Microbiol. Res. J.* 1(3):70-78.
- Hakkim F, Arivazhagan G, Boopathy R (2008). Antioxidant property of selected *Ocimum* species and their secondary metabolite content. *J. Med. Plant. Res.* 2(9):250-257.
- Kashyap CP, Ranjeet K, Vikrant A, Vipin K (2011). Therapeutic Potency of *Ocimum Kilimandscharicum* Guerke - A Review. *Global J. Pharmacol.* 5 (3):191-200.
- Kekuda TR (2012). *In vitro* Antibacterial Efficacy of Selected Plant Extracts, Streptomycin and their Combination. *Asian J. Res. Chem.* 5(6):791-793.
- Nakamura V, Ueda-Nakamura T, Bando E, Melo N, Cortez G, Filho D. Antibacterial (1999). Activity of *Ocimum gratissimum* L. Essential Oil. *Mem. Inst. Oswaldo Cruz.* 94(5):675-678.
- Nascimento GF, Locatelli J, Freitas PC, Silva GL (2000). Antibacterial activity of plant extracts and phytochemicals on antibiotic resistant bacteria. *Braz. J. Microbiol.* 31:247-256.
- Patil DD, Mhaske DK, Wadhawa GC (2011). Antibacterial and Antioxidant study of *Ocimum basilicum* Labiatae (sweet basil). *J. Adv. Pharm. Educ. Res.* 2:104-112.
- Prasannabalaji N, Muralitharan G, Sivanandan RN, Kumaran S, Pugazhvendan SR (2012). Antibacterial activities of some Indian traditional plant extracts. *Asian Pacific. J. Trop. Dis.* pp S291-S295.
- Sneha G, Margaret NJ, Sastry P, Jyothi Ch (2011). Evaluation of

- Antibacterial Activity of Ocimum. *Int. J. Pharmacogn. Phytochem. Res.* 3(3):89-92.
- Tchoumboungang F, Zollo PHA, Avlessi F, Alitonou GA, Sohounhloue DK, Ouamba JM, Tsomambet A, Okemy-Andissa N, Dagne E, Agnani H, Bessi re JM, Menut C (2014). Variability in the Chemical Compositions of the Essential Oils of Five Ocimum Species from Tropical African Area. *J. Essent. Oil Res.* 18:194-199.
- Wiat C (2006). *Medicinal Plants of the Asia-Pacific Drugs for the Future?* World Scientific Publishing Co. Pte. Ltd., New Jersey London Singapore Beijing Shanghai Hong Kong Taipei Chennai.
- Yilmaz MT (2012). Minimum inhibitory and minimum bactericidal concentrations of boron compounds against several bacterial strains. *Turk. J. Med. Sci.* 42 (2):1423-1429.
- Zuzarte M, Gonalves MJ, Canhoto J, Salgueiro L (2011). Antidermatophytic activity of essential oils. *Science against microbial pathogens: communicating current research and technological advances* pp. 1167-1178.

Full Length Research Paper

Chemical composition of essential oils from the stem barks of *Croton conduplicatus* (Euphorbiaceae) native to the Caatinga biome

Jackson Roberto Guedes da Silva Almeida^{1*}, Ana Valéria Vieira de Souza², Ana Paula de Oliveira¹, Uiliane Soares dos Santos², Maziele Dias de Souza², Luma dos Passos Bispo², Izabel Cristina Casanova Turatti³ and Norberto Peporine Lopes³

¹Núcleo de Estudos e Pesquisas de Plantas Medicinais (NEPLAME), Universidade Federal do Vale do São Francisco, 56.304-205, Petrolina, Pernambuco, Brazil.

²Empresa Brasileira de Pesquisa Agropecuária (EMBRAPA), 56.302-970, Petrolina, Pernambuco, Brazil.

³Departamento de Física e Química, Faculdade de Ciências Farmacêuticas de Ribeirão Preto, Universidade de São Paulo, 14.040-903, Ribeirão Preto, São Paulo, Brazil.

Received 8 March, 2014; Accepted 30 January, 2015

***Croton conduplicatus* is a species popularly known in the Brazilian Caatinga (semi-arid vegetation) as “quebra faca”. Essential oils from the stem barks of *C. conduplicatus* were obtained by hydrodistillation after 2, 3 and 4 h of extraction with a very high percentage (95.93, 96.69 and 98.45%, respectively) of identified total components present in crude essential oils. Analyses were made by gas chromatography/mass spectrometry (GC/MS). The most abundant constituents were α -pinene, β -pinene, camphor and (*E*)-caryophyllene. The occurrence of α -pinene and β -pinene has been reported in essential oils of several other species of *Croton*, indicating that this species is typically of the Euphorbiaceae family.**

Key words: *Croton conduplicatus*, essential oil, volatile constituents, medicinal plants, Caatinga.

INTRODUCTION

Croton (Euphorbiaceae) is one of the largest genera of flowering plants, with nearly 1300 species of herbs, shrubs and trees that are ecologically prominent and often important elements of secondary vegetation in the tropics and subtropics worldwide (Simionatto et al., 2007).

Some species of the genus *Croton*, such as *Croton cajucara*, *Croton zambesicus*, *Croton nepetaefolius* and *Croton celtidifolius*, have been described as medicinal

plants with their biological activities assessed. Amongst such plants studied to date, many have been revealed to display multiple biological activities, such as anti-inflammatory, antioxidant, antinociceptive, anticonvulsant and anxiolytic activities (Zhao et al., 2012).

Plants belonging to the genus *Croton* are well known for producing a variety of diterpenoids including pimarane, kaurane, labdane, cembrane, cleisthantane, and clerodane diterpenoids, with a wide range of

*Corresponding author. E-mail: jackson.guedes@univasf.edu.br. Tel/Fax: +55 (87) 21016862.

Author(s) agree that this article remain permanently open access under the terms of the [Creative Commons Attribution License 4.0 International License](https://creativecommons.org/licenses/by/4.0/)

Table 1. Chemical composition of essential oils of stem bark of *Croton conduplicatus* subjected to hydrodistillation for 2, 3 and 4 h of extraction.

Order	Compounds	RI*	GC/MS (%)		
			After 2 h	After 3 h	After 4 h
1	Tricyclene	922	0.39	0.78	0.50
2	α -Thujene	924	0.75	0.55	0.13
3	α -Pinene	932	32.87	35.35	25.84
4	Camphene	948	4.01	8.01	5.66
5	Sabinene	971	0.06	---	---
6	β -Pinene	977	13.56	16.77	7.79
7	Myrcene	988	0.72	0.50	0.24
8	<i>p</i> -Cymene	1024	0.72	0.56	0.44
9	Limonene	1028	1.24	1.40	0.72
10	1,8-Cineole	1031	1.46	0.29	0.54
11	γ -Terpinene	1056	0.15	0.21	0.14
12	<i>cis</i> -Linalool oxide	1069	0.13	---	---
13	<i>trans</i> -Linalool oxide	1086	0.56	0.81	0.36
14	Linalool	1100	2.18	1.24	0.86
15	NI	1104	0.24	---	---
16	<i>trans</i> -Pinocarveol	1140	---	0.31	---
17	Camphor	1146	7.30	9.32	6.37
18	Camphene hydrate	1154	---	0.19	0.15
19	Borneol	1171	0.33	0.33	0.64
20	Terpinen-4-ol	1179	0.87	0.52	0.29
21	α -Terpineol	1194	1.40	0.87	0.70
22	Thymol methyl ether	1228	0.70	---	0.49
23	Cyclosativene	1365	0.66	0.63	0.46
24	α -Copaene	1373	0.62	1.14	0.68
25	β -Elemene	1387	0.30	0.16	0.25
26	α -Gurjunene	1399	---	---	0.28
27	(<i>E</i>)-Caryophyllene	1416	7.80	4.66	5.07
28	β -Cubebene	1426	0.64	---	0.34
29	NI	1445	---	---	0.33
30	α -Humulene	1452	1.47	0.84	1.18
31	NI	1464	---	---	0.51
32	γ -Muurolene	1472	2.08	0.19	2.76
33	Germacrene D	1477	0.36	---	0.80
34	α -Selinene	1492	---	---	0.37
35	α -Muurolene	1496	3.66	3.21	1.79
36	β -Bisabolene	1506	---	0.30	0.27
37	δ -Amorphene	1509	---	---	0.38
38	γ -Cadinene	1509	0.23	---	---
39	NI	1511	---	---	0.44
40	δ -Cadinene	1515	0.31	0.26	1.78
41	NI	1538	0.18	---	---
42	Elemol	1545	0.30	---	---
43	Hedycariol	1545	---	---	0.57
44	Germacrene B	1554	---	0.20	0.32
45	Caryophyllene oxide	1578	6.63	3.88	6.91
46	Viridiflorol	1585	---	---	0.22
47	Guaiol	1593	0.66	0.41	0.83
48	Globulol	1600	---	---	1.12
49	Humulene epoxide II	1605	0.84	0.50	1.16

Table 1. Cont'd.

50	NI	1622	2.16	1.70	---
51	Aristolene	1629	---	1.00	1.32
52	NI	1633	0.39	0.48	---
53	Hinesol	1636	---	0.76	0.76
54	NI	1652	0.65	0.63	---
55	NI	1667	0.45	0.50	---
Total		-	95.93	96.69	98.45

*RI: Retention indices on DB-5MS column (relative to *n*-alkanes); NI: not identified compound; (---): Not detected.

biological activities (Pudhom et al., 2007). On the other hand, essential oils are other important class of secondary metabolites in this genus. Recently, Salatino et al. (2007) reported the study of the essential oils of about thirty species of *Croton*. The results indicated that some of these oils are rich in terpenoids and phenylpropanoids, and others are rich only in terpenoids (Salatino et al., 2007).

Despite of the large array of data on other *Croton* species, the knowledge about *Croton conduplicatus* is scarce. This species is popularly known in the Brazilian Caatinga as "quebra faca". Decoction of its leaves and stem barks are used in folk medicine to treat influenza, headache, indigestion, stomach problems and stomachache (Cartaxo et al., 2010). To the best of our knowledge, no phytochemical and pharmacological studies have previously been reported on this species.

This paper presents for the first time the chemical composition of *C. conduplicatus* stem barks essential oils by gas chromatography/mass spectrometry (GC/MS).

MATERIALS AND METHODS

Plant

Stem barks of *C. conduplicatus* Kunth. were collected from a single individual in September 2012 in Petrolina (Coordinates: S 09°03'54"; W 40°19'12"), State of Pernambuco, Brazil. A voucher specimen (HTSA2421) was deposited at the Herbário do Trópico Semiárido (HTSA) of the Empresa Brasileira de Pesquisa Agropecuária (EMBRAPA). Considering that the plant is a shrub, the stem barks were cut close to the ground.

Extraction of essential oils

The fresh stem barks (100 g) were cut into pieces, and subjected to hydrodistillation for 2, 3 and 4 h in a modified Clevenger-type apparatus. The oils were dried over anhydrous sodium sulfate. The essential oils obtained have yellow color and characteristic odor. The oils were stored in a refrigerator until the analysis by GC/MS.

Analysis of essential oils

The substances present in the essential oil of *C. conduplicatus* were investigated on a Shimadzu QP-2010 gas chromatograph

interfaced to a mass spectrometer (GC/MS). The following conditions were used: DB-5MS column Agilent Technologies (30 m × 0.25 mm × 0.25 μm); helium (99.999%) carrier gas at a constant flow of 1.1 ml/min; 1.0 μl injection volume; injector split ratio of 1:10; injector temperature 250°C; electron impact mode at 70 eV; ion-source temperature 280°C and transfer line temperature 260°C. The oven temperature was programmed from 60°C, with an increase of 3°C min⁻¹ to 240°C.

A mixture of linear hydrocarbons (C₉H₂₀–C₂₁H₄₀) was injected under the same experimental conditions as samples, and identification of the constituents was performed by comparing the spectra obtained with those of the equipment database (Wiley 7 lib and Nist 08 lib) and by using the Kovats Index, each constituent was calculated as previously described (Adams, 1995; Van den Dool and Kratz, 1963). The data were acquired and processed with a PC with Shimadzu GC/MS Solution software.

RESULTS AND DISCUSSION

In every extraction, 100 g of *C. conduplicatus* stem barks were used and the crude oils yield was found to be 0.90, 0.97 and 0.97 ml, for 2, 3 and 4 h of extraction, respectively. The CG/MS analysis led to the identification of 95.93, 96.69 and 98.45% of the total components present in crude essential oils.

The chemical constituents of the essential oil of *C. conduplicatus* were identified by comparing their mass spectral data with reference spectra in the computer library. The identified compounds are as shown in Table 1 according to their retention indexes.

The main compounds found in the oil of the stem barks after 2, 3 and 4 h of extraction were α-pinene (32.87, 35.35 and 25.84%, respectively), β-pinene (13.56, 16.77 and 7.79%, respectively), camphor (7.30, 9.32 and 6.37%, respectively) and (*E*)-caryophyllene (7.80, 4.66 and 5.07%, respectively). Variation in extraction time was performed to verify their influence on the yield and the chemical composition of the essential oil. Depending on the compound of interest, the time of extraction can be adjusted.

In light of these chemical evidences, some authors purpose to consider that the co-occurrence of α and β-pinene might be a characteristic of the genus *Croton*, however, in the light of a larger number thus far studied, taxa, β-caryophyllene and linalool seem to be equally a

frequent major constituents of many *Croton* spp. (Radulovic et al., 2006).

Particular relevance should also be given to the presence of minor, but not negligible compounds detected in our samples as caryophyllene oxide, camphene and α -muurolene. The relatively high content of camphene in the stem oil differentiates *Croton decaryi* from the other members of the genus *Croton* since there are no published data on a *Croton* spp. containing this monoterpene hydrocarbon as one of the major constituents (Radulovic et al., 2006). Thus, camphene could be considered as chemotaxonomic marker for *C. decaryi*.

Conclusion

C. conduplicatus has been examined for the first time for the essential oil obtained by hydrodistillation of fresh stem barks. GC/MS has been provided in order to chemically characterize the essential oil, evidencing a monoterpene prevalence. In comparison to the others essential oils from *Croton* spp., the oil of this species shows constituents that are present in other species of this genus.

ACKNOWLEDGEMENT

This work was supported by grants from Brazilian agency CNPq (Process 151322/2013-9).

Conflict of interest

Authors declare that there are no conflicts of interest.

REFERENCES

- Adams RP (1995). Identification of essential oil components by Gas Chromatography/Mass Spectroscopy. 4. ed. USA: Allured Publishing Corporation Carol Stream, IL.
- Cartaxo SL, Souza MMA, Albuquerque UP (2010). Medicinal plants with bioprospecting potential used in semi-arid northeastern Brazil. *J. Ethnopharmacol.* 131(2):326-342.
- Pudhom K, Vilaivan T, Ngamrojanavanich N, Dechangvipart S, Sommit D, Petsom A, Roengsumran S (2007). Furanocembranoids from the stem bark of *Croton oblongifolius*. *J. Nat. Prod.* 70(4):659-661.
- Radulovic N, Mananjaraso E, Harinantenaina L, Yoshinori A (2006). Essential oil composition of four *Croton* from Madagascar and their chemotaxonomy. *Biochem. Syst. Ecol.* 34(8):648-653.
- Salatino A, Salatino MLF, Negri G (2007). Traditional uses, chemistry and pharmacology of *Croton* species (Euphorbiaceae). *J. Braz. Chem. Soc.* 18(1):11-33.
- Simionatto E, Bonani VFL, Morel AF, Poppi NR, Raposo-Junior JL, Stuker, CZ, Peruzzo, GM, Peres MTL, Hess SC (2007). Chemical composition and evaluation of antibacterial and antioxidant activities of the essential oil of *Croton urucurana* Baillon (Euphorbiaceae) stem bark. *J. Braz. Chem. Soc.* 18(5):879-885.
- Van den Dool H, Kratz PDJA (1963). Generalization of the retention index system including linear temperature programmed gas-liquid partition chromatography. *J. Chromatogr.* 11:463-471.
- Zhao J, Fang F, Yu L, Wang G, Yang L (2012). Anti-nociceptive and anti-inflammatory effects of *Croton crassifolius* ethanol extract. *J. Ethnopharmacol.* 142(2):367-373.



African Journal of Pharmacy and Pharmacology

Related Journals Published by Academic Journals

- *Journal of Medicinal Plant Research*
- *African Journal of Pharmacy and Pharmacology*
- *Journal of Dentistry and Oral Hygiene*
- *International Journal of Nursing and Midwifery*
- *Journal of Parasitology and Vector Biology*
- *Journal of Pharmacognosy and Phytotherapy*
- *Journal of Toxicology and Environmental Health Sciences*

academicJournals